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WITNESS my hand this Eighteenth day of August 2003

LEANNE MYNOTT

MANAGER EXAMINATION SUPPORT

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### PROVISIONAL SPECIFICATION

for the invention entitled:

"Diagnostic and therapeutic agents"

The invention is described in the following statement:

### DIAGNOSTIC AND THERAPEUTIC AGENTS

#### FIELD OF THE INVENTION

The present invention relates generally to diagnostic and therapeutic agents. More particularly, the present invention provides mammalian transcription factors which function in the modulation of expression of genetic sequences. The present invention further provides nucleic acid molecules encoding the transcription factors as well as nucleic acid and/or proteinaceous molecules with which the transcription factors interact.

The transcription factors of the present invention or molecules interacting with same may be used *inter alia* in the generation of a range of diagnostic and therapeutic agents for a range of conditions.

#### BACKGROUND OF THE INVENTION

Reference to any prior art in this specification is not, and should not be taken as, an acknowledgment or any form of suggestion that this prior art forms part of the common

general knowledge in any country.

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20 Bibliographic details of references provided in the subject specification are listed at the end of the specification.

The increasing sophistication of recombinant DNA techniques has provided significant progress in understanding the mechanisms involved in regulating eukaryotic gene expression. This is greatly facilitating research and development in the plant, agricultural, medical and veterinary industries. Transcription factors are an important component in the control of gene expression. However, despite their importance, mammalian transcription factors have not been well investigated for their diagnostic and therapeutic potential.

30 RNA polymerases in eukaryotic cells cannot initiate transcription alone; before transcription can begin, they require interaction between transcription factors and the

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promoter. These factors assemble at the promoter and, via a series of steps, facilitate both the binding of RNA polymerase II to the promoter and its subsequent phosphorylation and release to initiate transcription.

In addition to these general transcription factors, many thousands of transcription activators and/or negative regulators (inhibitors) exist, which control the process of initiation of gene transcription from great distances along the DNA. These factors influence the timing and extent of transcription of a particular gene. Indeed, they control whether and to what extent a particular gene is transcribed in a cell of a particular tissue type. Although most gene regulators identified to date have been found to be proteins, some transcription factors may also be RNA molecules.

In Drosophila, the transcription factor known as "Grainyhead" regulates key developmental process in the embryo and is encoded by the gene grainyhead. During development, Grainyhead is initially involved in dorsal/ventral and terminal patterning of the newly fertilized embryo through the formation of multi-protein complexes that repress transcription from the decapentaplegic, tailless and zerknuellt genes (Huang et al., Genes Dev. 9: 3177-3189, 1995; Liaw et al., Genes Dev. 9: 3163-3176, 1995). Later, grainyhead is predominantly expressed in the embryonic central nervous system in cuticle-producing tissues, where it binds to promoters and influences transcription from other developmentally regulated genes including engrailed, fushi tarazu and Ultrabithorax (Bray et al., Genes Dev. 3: 1130-1145, 1989; Dynlacht et al., Genes Dev. 3: 1677-1688, 1989; Biggin and Tjian, Cell 53: 699-711, 1988; Soeller et al., Genes Dev. 2: 68-81, 1988; Dynlacht et al., Cell 56: 563-576, 1991; Attardi and Tjian, Genes Dev. 7: 1341-1353, 1993; Uv et al., Mol. Cell Biol. 14: 4020-4031, 1994).

The importance of grainyhead in Drosophila development is emphasised by the embryonic lethal phenotype observed in flies carrying mutations in this gene. The embryos have flimsy cuticles, grainy and discontinuous head skeletons and patchy tracheal tubes (Bray and Kafatos, Genes Dev. 5: 1672-1683, 1991). A neuroblast-specific isoform of the protein, arising from alternate splicing, has also been identified. A mutation that abolishes

this isoform is pupal- and adult- lethal, and flies demonstrate uncoordinated movements (Uv et al., Mol. Cell Biol. 17: 6727-6735, 1997).

Mammalian homologs of grainyhead have previously been proposed, including three genes designated CP2, LBP-1a and LBP-9. Studies have implicated them in a wide variety of cellular and developmental events including T cell proliferation, globin gene expression and steroid biosynthesis (Sueyoshi et al., Mol. Cell Biol. 15: 4158-4166, 1995; Jane et al., EMBO J. 14: 97-105, 1995; Volker et al., Genes Development 11: 1435-1446, 1997; Zhou et al., Mol. Cell Biol. 20: 7662-7672, 2000). However, in situ analyses of both CP2 and LBP-1a reveal ubiquitous expression of both genes, unlike the highly restricted pattern observed with grainyhead in Drosophila (Bray et al., 1989, supra; Dynlacht et al., 1989, supra; Bray and Kafatos, 1991, supra; Ramamurthy et al., J. Biol. Chem. 276: 7836-7842, 2001). It is concluded, therefore, that these genes are not close homologs of grainyhead.

15 There is a need to identify other mammalian transcription factors and in particular close mammalian homologs of Grainyhead and to use these to develop a range of diagnostic and therapeutic agents.

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#### SUMMARY OF THE INVENTION

Throughout this specification, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the 5 inclusion of a stated element or integer or group of elements or integers but not the exclusion of any other element or integer or group of elements or integers.

Nucleotide and amino acid sequences are referred to by a sequence identifier number (SEQ ID NO:). The SEQ ID NOs: correspond numerically to the sequence identifiers <400>1 10 '(SEQ ID NO:1), <400>2 (SEQ ID NO:2), etc. A sequence listing is provided at the end of the specification. A summary of the SEQ ID NOs is provided in Table 1.

Genetic sequences were studied exhibited homology at the nucleotide and/or amino acid level to a Drosophila gene, the product of which is involved in body patterning where a fine balance between activation and inhibition of gene expression is critical to the correct development of cells and tissues into functional organisms. A large number of different families of transcription factors play a critical role in ensuring that this balance is maintained during embryological development. One such transcription factor, cloned from Drosophila and well-characterized, is Grainyhead (hereinafter referred to by its 20 abbreviation, GRH). GRH is encoded by the gene grainyhead (grh). The inventors observed that the identity of previously published putative grh mammalian homologs showed much more ubiquitous expression compared with the highly restricted pattern exhibited by Drosophila grh. Furthermore, sequence similarity between the proposed mammalian homologs and the Drosophila grh sequence was relatively low. In accordance with the present invention, true grh homologs were identified and derived from mammalian tissue such as human and mouse tissue.

Accordingly, one aspect of the present invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides encoding or complementary to a sequence encoding a mammalian homolog of Drosophila GRH. A mammalian homolog of GRH is 30 referred to herein as M-GRH. The corresponding gene is referred to as M-grh. A M-grh is

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deemed a homolog of *Drosophila grh* (*D-grh*). If it comprises a nucleotide sequence having 60% or greater similarity to the nucleotide sequence of *D-grh* after optimal alignment. Likewise, a M-GRH is so defined if it comprises an amino acid sequence having 60% or greater similarity to the amino acid sequence of *Drosophila* GDH (*D-GRH*). There are four isoforms of *Drosophila grh* designated *D-grh* P1, *D-grh* P2, *D-grh* P3 and *D-grh* P4. The nucleotide sequence encoding *D-grh* is set forth in SEQ ID NO:17 and SEQ ID NO:34, SEQ ID NO:36 and SEQ ID NO:38, respectively. Mammalian sequences encompassed by the present invention include those derived from tissues of mouse and human including, for example, mouse embryo, human fetal brain and placenta, and mouse and human kidney. Reference herein to *Drosophila grp* includes any or all of its isoforms P1-P4.

The mammalian sequences identified by the present inventors show higher percentages of similarity to the *D-grh* sequence than the already identified mammalian sequences designated *CP2*, *LBP-1a* and *LBP-9*. In accordance with the present invention, it is proposed that the M-grh homologs disclosed are "true" grh homologs relative to *CP2*, *LBP-1a* and *LBP-9*. As a result of the analysis herein described, it is shown that the earlier sequences align phylogenetically with another distinct *Drosophila* factor, designated *Drosophila CP2*. A new family of transcription factors, highly conserved from *Drosophila* to human and having distinct tissue-specificity profiles, is now described in accordance with the present invention.

The true M-grh homologs of the present invention include mammalian grainyhead (gene: mgr; expression product: MGR), brother of mgr (gene: bom; expression product: BOM) and sister of mgr (gene som: protein: SOM). MGR has multiple isoforms including MGR p49 and MGR p70 in humans and MGR p61 in mice. A summary of the SEQ ID NOs for the M-grh and M-GRH molecules of the present invention are shown in Table 2. The sequences are provided in the Sequence Listing.

30 The present invention provides, therefore, expression products of the M-grh genes, mgr, bom and som as well as derivatives and homologs thereof. This aspect of the present

invention does not extend to CP2, LBP-1a or LBP-9.

Accordingly, another aspect of the present invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides encoding a polypeptide comprising a predicted amino acid sequence substantially as set forth in SEQ ID NO:2 (human MGR p49), SEQ ID NO:4 (human MGR p70), SEQ ID NO:6 (human BOM), SEQ ID NO:8 (human SOM), SEQ ID NO:10 (murine MGR p49), SEQ ID NO:12 (murine MGR p70), SEQ ID NO:14 (murine BOM) or SEQ ID NO:16 (murine SOM) or an amino acid sequence having at least about 60% similarity to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16 after optimal alignment.

The preferred nucleic acid molecules comprise sequences of nucleotides substantially as set forth in SEQ ID NO:1 (human mgr p49), SEQ ID NO:3 (human mgr p70), SEQ·ID NO:5 (human bom), SEQ ID NO:7 (human som), SEQ ID NO:9 (murine mgr p61), SEQ ID NO:11 (murine mgr p70), SEQ ID NO:13 (murine bom) or SEQ ID NO:15 (murine som) or complementary forms thereof, or a nucleotide sequence having at least about 60% similarity to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 after optimal alignment or their complementary forms or a nucleotide sequence capable of hybridizing to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or complementary forms thereof under low stringency conditions. Again, this aspect of the present invention does not extend to nucleic acid molecules encoding CP2, LBP-1 and LBP-9.

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The present invention further extends to recombinant forms of the M-GRH molecules. Preferred recombinant M-GRH molecules having amino acid sequences defined in parenthesis include human MGR p49 (SEQ ID NO:2), human MGR p70 (SEQ ID NO:4), human BOM (SEQ ID NO:6), human SOM (SEQ ID NO:8), murine MGR p61 (SEQ ID NO:10), murine MGR p70 (SEQ ID NO:12), murine BOM (SEQ ID NO:14) and murine SOM (SEQ ID NO:16).

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Reference to "M-GRH" molecules include derivatives, homologs and analogs thereof.

The mammalian transcription factors of the present invention are proposed to be involved in the regulation of expression of a range of genes such as but not limited to developmentally regulated genes involved in determining patterning. Some of the genes regulated encode critical products, the absence or malfunctioning of which, is proposed to lead to unwanted phenotypes and/or predispositions to certain medical conditions. That is, the presence of a mutation in and/or malfunction of a M-grh including over or under expression of the transcription factors of the present invention are proposed to cause incorrect regulation of one or more of these genes thereby leading to an inappropriate phenotype. The ability to detect mutations in the nucleotide sequences encoding the M-grh homologs permits the detection of a range of abnormalities or a predisposition for development of abnormalities. Furthermore, as many of the genes will be developmentally regulated genes, identification of the transcription factors permits identification of unknown developmentally regulated genes.

Accordingly, another aspect of the present invention contemplates a method for detecting a variation in a polynucleotide sequence encoding a M-GRH transcription factor.

Furthermore, the isolated nucleic acid molecules of the present invention may be able to be used to correct such an abnormality in a subject in need thereof or at risk of developing an abnormality. The nucleic acid molecules of the present invention may be comprised, therefore, within a suitable vector for delivery of all or part of the sequence to a recipient cell or tissue. The nucleic acid molecule or part thereof could also be administered directly for transient expression. The present invention provides, therefore, the potential for both a diagnostic and a therapeutic capability.

Accordingly, a further aspect of the present invention contemplates a genetic construct comprising a nucleotide sequence selected from SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15 or a

nucleotide sequence having at least 60% similarity to one or more of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 after optimal alignment or a nucleotide sequence capable of hybridizing to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or a complementary form thereof under low stringency conditions.

In a related embodiment, the present invention provides a genetic construct comprising a promoter or functional equivalent thereof operably linked to a nucleotide sequence of the invention.

Genes are represented herein in lower case italics. Expression products (e.g. proteins or RNA) are represented in upper case, non-itallic letters. A summary of the genes and their expression products is provided in Table 1.

TABLE 1
Abbreviations

GENE	EXPRESSION PRODUCT	
grainyhead (grh)	Grainyhead (GRH)	
mammalian grainyhead homologs (M-grh)	mammalian grainyhead homologs (M-GRH)	
mammalian grainyhead (mgr)	mammalian Grainyhead (MGR)	
brother of mammalian grainyhead (bom)	brother of mammalian grainyhead (BOM)	
sister of mammalian grainyhead (som)	sister of mammalian grainyhead (SOM)	

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A summary of sequence identifiers used throughout the specification is Table 2.

TABLE 2
Summary of sequence identifiers

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SEQUENCE	NAME	DESCRIPTION
ID NO.		
1	human <i>mgr</i> p49	Nucleotide sequence encoding mammalian
		grainyhead derived from human fetal brain
2	human MGR p49	Predicted amino acid sequence corresponding to SEQ ID NO:1
3	human <i>mgr</i> p70	Nucleotide sequence encoding mammalian grainyhead being an isoform of SEQ ID NO:1, derived from human kidney
4	human MGR p70	Predicted amino acid sequence corresponding to SEQ ID NO:3
5	human bom	Nucleotide sequence encoding mammalian grainyhead derived from human placenta
6	human BOM	Predicted amino acid sequence corresponding to SEQ ID NO:5
7	human som	Nucleotide sequence encoding mammalian grainyhead
8	human SOM	Predicted amino acid sequence corresponding to SEQ ID NO:7
9	murine <i>mgr</i> p61	Nucleotide sequence encoding mammalian grainyhead derived from 17.5 day murine embryo
10	murine MGR p61	Predicted amino acid sequence corresponding to SEQ ID NO:9
11	murine mgr p70	Nucleotide sequence encoding mammalian grainyhead being an isoform of SEQ ID NO:9, derived from murine kidney
12	murine MGR p70	Predicted amino acid sequence corresponding to SEQ ID NO:11
13	murine bom	Nucleotide sequence encoding mammalian grainyhead derived from a murine embryonic carcinoma cell line (p19)
14	murine BOM	Predicted amino acid sequence corresponding to SEQ ID NO:13
15	murine som	Nucleotide sequence encoding mammalian grainyhead
16	murine SOM	Predicted amino acid sequence corresponding to SEQ ID NO:15
17	grh-P1	Nucleotide sequence encoding the Drosophila

SEQUENCE	NAME	DESCRIPTION
ID NO.		transcription factor designated Grainyhead (grh)
18	GRH-P1	Amino acid sequence corresponding to SEQ ID NO:18
19-20	human p49 mgr	primers
21-22	human p70 mgr	primers
23-24	human <i>bom</i>	primers
25-26	murine p70 mgr	primers
27-28	murine p61 mgr	primers
29-30	murine bom	primers
31-32	human S14	primers
33	Drosophila dopa decarboxylase	promoter
34	Drosophila PCNA	promoter
35	human Engrailed-1	promoter
36	grh-P2	Nucleotide sequence encoding the <i>Drosophila</i> transcription factor designated <i>Grainyhead</i> (grh) isoform P2
37	GRH-P2	Amino acid sequence corresponding to SEQ ID NO:36
38	grh-P3	Nucleotide sequence encoding the Drosophila transcription factor designated Grainyhead (grh) isoform P3
39	GRH-P3	Amino acid sequence corresponding to SEQ ID NO:38
40	grh-P4	Nucleotide sequence encoding the <i>Drosophila</i> transcription factor designated <i>Grainyhead</i> (grh) isoform P4
41	GRH-P4	Amino acid sequence corresponding to SEQ ID NO:40

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#### BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is a representation showing that mgr genomic locus encodes two distinct isoforms. (A) Alignment of the predicted NH2-terminal amino acid sequence of the p70 5 isoform of MGR and BOM. Amino acid identity is denoted by shared upper case letters and similarity by the (+) symbol. The first amino acids shared between p61 MGR and p70 MGR are given in bold. (B) Structure of the human and murine mgr genomic loci. Human genomic sequence was downloaded from the GenBank database (Accession Number AC010969) and aligned with cDNA sequences. Murine genomic clones were obtained from a 129 library and mapped by Southern analysis and PCR. Exons are denoted as E1-8 in human and E1-9 in murine. The two human MGR isoforms are denoted as p70 and p49 MGR and the two murine isoforms as p70 and p61 MGR. The scale of 1 kb is shown. (C) Identification of the murine p61 MGR promoter. Sequence was obtained from intron three from the MGR genomic locus and analyzed using the weight matrices of Bucher, J. Mol. Biol. 212: 563-578, 1990. The CAP site, TATA box and GC box are indicated. The cDNA start site is shown in arrows, the first ATG is given in bold and the splice site at the end of the first exon of p61 MGR is indicated.

Figure 2 is a photographic representation showing that p70 MGR binds to Drosophila gene regulatory sequences which bind grh. (A) p70 MGR binds to the Drosophila PCNA promoter. Nuclear extract from the JEG-3 cell line was studied in an EMSA with a PCNA promoter probe in the presence and absence of anti-MGR specific antisera. Antisera 611 was raised against peptides common to the p70 and p49 MGR proteins in the dimerization domain and antisera 67 was raised against unique peptides in the NH2-terminal domain of p70 MGR. The migration of the MGR complex is shown in arrows. (B) p70 MGR binds to the Drosophila dopo decarboxylase promoter. Experimental conditions were as described for (A).

Figure 3 are representations showing that p70 MGR binds to and transactivates the human En-1 promoter. (A) Identification of a grh consensus DNA binding site in the human En-1 promoter. The consensus sequence for grh DNA binding compiled from an alignment of

the Drosophila Ultrabithorax, Dopa decarboxylase and fushi tarazu promoters was compared with the sequence of the proximal human En-1 promoter and the Drosophila engrailed promoter. The closed bracket indicates the extend of the grainyhead binding site in the engrailed promoter as defined by DNAseI footprinting. (B) Human p70 MGR binds to the human En-1 promoter. Nuclear extract from the JEG-3 cell line was studied in an EMSA with a Ddc promoter probe in the presence of pre-immune sera (lane 1), anti-MGR specific antisera 67 (detailed in legend to Figure 2) (lane 2) or cold competitor DNA (lanes 3-5). A 50-fold excess of the Ddc probe was used in lane 3 and a 10- and 20-fold excess of a human En-1 promoter probe in lanes 4 and 5, respectively. The migration of the MGR/DNA complex is shown by arrows. (C) Human p70 MGR transactivates the En-1 promoter. COS cells were transiently transfected with the proximal En-1 promoter containing the MGR binding site linked to a mimimal y-globin promoter and a firefly luciferase reporter gene (solid columns), the miminal γ-globin promoter/luciferase reporter gene (open columns) and the TK promoter linked to the Renilla luciferase reporter gene (hatched columns) in the presence and absence of a p70 MGR expression vector (PCI-p70 MGR) as indicated. Transfection with the empty vector (pCI) served as the control. Luciferase levels were corrected for protein concentration and values were derived from two independent experiments performed in triplicate.

# DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is predicated in part on the identification of mammalian homologs of the *Drosophila* transcription factor known as Grainyhead (GRH). GRH is encoded by the gene, grainyhead (grh) In *Drosophila*, mutations in this gene are associated with embryonic lethal phenotypes, indicating the importance of the gene for normal development and function. The mammalian homologs are proposed to be involved in the regulation of developmental and/or non-developmental genes. Identification and isolation of the mammalian homologs of grh (M-grh) enable the development of a range of diagnostic and therapeutic agents useful in the detection and treatment of genetic disorders.

The present invention provides, therefore, a family of mammalian-derived transcription factors, highly related from Drosophila to mammals. These transcription factors are more highly conserved than CP2, LBP-1a and LBP-9. The present invention does not extend to CP2, LBP-1 and LBP-9. Reference to a mammal in this context includes a human, livestock animal (e.g. sheep, cow, horse, pig, donkey, goat), laboratory test animal (e.g. mouse, rat, rabbit, guinea pig), companion animal (e.g. dog, cat) or captive wild animal. Most preferably, the animal is a human or murine species. Sources of the isolated nucleic acid molecules include a range of tissues, such as mouse embryo, human fetal brain and placenta, and mouse and human kidney. In view of the highly conserved nature of this family of M-grh nucleotide sequences, however, corresponding homologs from other tissues and from other mammalian species are intended to be included within the scope of the present invention. The term "homolog" as used herein, therefore, extends to encompass transcription factors from mammalian species encoded by nucleotide sequences which have substantial similarity to Drosophilia grh or a conserved region thereof. At the protein level, a homolog includes an amino acid sequence and/or tertiary structure having similarity to Drosophila GRH. In cases where the expression product of the M-grh is RNA, a homolog is defined by reference to the similar ribonucleotide sequence to that encoded by Drosophila grh.

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M-ghd or M-GRH, i.e. a mammalian homolog of Drosophila grh or GRH is defined as

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such by having a nucleotide or amino acid sequence which has 60% or greater similarity after optimal alignment to *Drosophila grh* or GRH.

Accordingly, one aspect of the present invention provides an isolated nucleic acid molecule comprising a sequence of nucleotides encoding or complementary to a sequence encoding a mammalian homolog of *Drosophila grh*.

Reference to a mammalian homolog of *Drosophila* GRH (i.e. a M-GRH) preferably includes the mammalian homolog of grainyhead (MGR), brother of MGR (BOM) and sister of MGR (SOM). These transcription factors are encoded by *mgr*, *bom* and *som*, respectively. Reference to "MGR", "BOM" and "SOM" or *mgr*, *bom* and *som* includes all mutants, derivatives, homologs and analogs thereof. The present invention further extends, however, to all novel mammalian homologs of *Drosophila grh* but does not encompass CP2, LBP-1a or LBP-9. The nucleotide sequences for *Drosophila grh* are set forth in SEQ ID NO:17, SEQ ID NO:34, SEQ ID NO:36 and SEQ ID NO:38, respectively. Consequently, a mammalian homolog is defined herein as comprising a nucleotide sequence having at least about 60% sequence similarity to SEQ ID NO:17 or SEQ ID NO:36 or SEQ ID NO:36 or SEQ ID NO:38 after optimal alignment and/or being capable of hybridizing to SEQ ID NO:17 or SEQ ID NO:34 or SEQ ID NO:36 or SEQ ID NO:38 or its complementary form under low stringency conditions.

Accordingly, another aspect of the present invention provides an isolated nucleic acid molecule encoding a mammalian transcription factor or a functional part thereof comprising a sequence of nucleotides having at least 60% similarity to SEQ ID NO:17 or SEQ ID NO:34 or SEQ ID NO:36 or SEQ ID NO:38 after optimal alignment and/or being capable of hybridizing to SEQ ID NO:17 or its complementary form under low stringency conditions.

In a preferred embodiment, the isolated nucleic acid molecule encodes a proteinaceous form of a transcription factor. Examples of such mammalian protein transcription factors include human MGR p49 (SEQ ID NO:2), human MGR p70 (SEQ ID NO:4), human

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BOM (SEQ ID NO:6), human SOM (SEQ ID NO:7), murine MGR p61 (SEQ ID NO:10), murine MGR p70 (SEQ ID NO:12), murine BOM (SEQ ID NO:14) and murine SOM (SEQ ID NO:16).

Accordingly, another aspect of the present invention is directed to an isolated nucleic acid molecule comprising a sequence of nucleotides encoding a polypeptide having transcription factor activity and comprising an amino acid sequence substantially as set forth in SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14 or SEQ ID NO:16 or an amino acid sequence having at least about 60% similarity to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14 or SEQ ID NO:16 after optimal alignment wherein said polypeptide is a mammalian homolog of *Drosophia* GRH.

Such a polypeptide is referred to herein as a M-GRH.

Preferred percentage amino acid similarity levels include at least about 61% or at least about 62% or at least about 63% or at least about 64% or at least about 65% or at least about 66% or at least about 67% or at least about 68% or at least about 69% or at least about 70% or at least about 71% or at least about 72% or at least about 73% or at least about 74% or at least about 75% or at least about 76% or at least about 77% or at least about 78% or at least about 80% or at least about 81% or at least about 82% or at least about 83% or at least about 84% or at least about 85% or at least about 86% or at least about 87% or at least about 88% or at least about 89% or at least about 90% or at least about 91% or at least about 92% or at least about 93% or at least

about 94% or at least about 95% or at least about 96% or at least about 97% or at least about 98% or at least about 99% similarity.

This aspect of the present invention includes derivatives of M-GRH molecules. Such derivatives include non-active fragments which encompass *inter alia* the binding domain as well as active isoforms.

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A "derivative" of a polypeptide of the present invention also encompasses a portion or a part of a full-length parent polypeptide, which retains the transcription factor activity of the parent polypeptide. Such "biologically-active fragments" include deletion mutants and small peptides, for example, of at least 10, preferably at least 20 and more preferably at least 30 contiguous amino acids, which exhibit the requisite activity. Peptides of this type may be obtained through the application of standard recombinant nucleic acid techniques or synthesized using conventional liquid or solid phase synthesis techniques. For example, reference may be made to solution synthesis or solid phase synthesis as described, for example, in Chapter 9 entitled "Peptide Synthesis" by Atherton and Shephard which is included in a publication entitled "Synthetic Vaccines" edited by Nicholson and published by Blackwell Scientific Publications. Alternatively, peptides can be produced by digestion of an amino acid sequence of the invention with proteinases such as endoLys-C, endoArg-C, endoGlu-C and staphylococcus V8-protease. The digested fragments can be purified by, for example, high performance liquid chromatographic (HPLC) techniques. Any such fragment, irrespective of its means of generation, is to be understood as being encompassed by the term "derivative" as used herein.

In another embodiment, the present invention provides an isolated nucleic acid molecule encoding a mammalian transcription factor homolog of *Drosophila grh* (i.e. a M-GRH) and comprising a nucleotide sequence selected from SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 and SEQ ID NO:15 or a nucleotide sequence having at least about 60% similarity to any one of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 after optimal alignment or a nucleotide sequence capable of hybridizing to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:13 or SEQ ID NO:15 or a complementary form thereof under low stringency conditions.

Preferably, percentage nucleotide similarity levels include at least about 61% 61% or at least about 62% or at least about 63% or at least about 64% or at least about 65% or at least about 66% or at least about 69% or at

least about 70% or at least about 71% or at least about 72% or at least about 73% or at least about 74% or at least about 75% or at least about 76% or at least about 77% or at least about 78% or at least about 80% or at least about 81% or at least about 82% or at least about 83% or at least about 84% or at least about 85% or at least about 86% or at least about 87% or at least about 88% or at least about 89% or at least about 90% or at least about 91% or at least about 92% or at least about 93% or at least about 94% or at least about 95% or at least about 96% or at least about 97% or at least about 98% or at least about 99% similarity.

- 10 The term "similarity" as used herein includes exact identity between compared sequences at the nucleotide or amino acid level. Where there is non-identity at the nucleotide level, "similarity" includes differences between sequences which result in different amino acids that are nevertheless related to each other at the structural, functional, biochemical and/or conformational levels. Where there is non-identity at the amino acid level, "similarity" includes amino acids that are nevertheless related to each other at the structural, functional, biochemical and/or conformational levels. In a particularly preferred embodiment, nucleotide and sequence comparisons are made at the level of identity rather than similarity.
- Terms used to describe sequence relationships between two or more polynucleotides or polypeptides include "reference sequence", "comparison window", "sequence similarity", "sequence identity", "percentage of sequence similarity", "percentage of sequence identity", "substantially similar" and "substantial identity". A "reference sequence" is at least 12 but frequently 15 to 18 and often at least 25 or above, such as 30 monomer units, inclusive of nucleotides and amino acid residues, in length. Because two polynucleotides may each comprise (1) a sequence (i.e. only a portion of the complete polynucleotide sequence) that is similar between the two polynucleotides, and (2) a sequence that is divergent between the two polynucleotides, sequence comparisons between two (or more) polynucleotides are typically performed by comparing sequences of the two polynucleotides over a "comparison window" to identify and compare local regions of sequence similarity. A "comparison window" refers to a conceptual segment of typically

12 contiguous residues that is compared to a reference sequence. The comparison window may comprise additions or deletions (i.e. gaps) of about 20% or less as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Optimal alignment of sequences for aligning a comparison window may be conducted by computerized implementations of algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Drive Madison, WI, USA) or by inspection and the best alignment (i.e. resulting in the highest percentage homology over the comparison window) generated by any of the various methods selected. Reference also may be made to the BLAST family of programs as for example disclosed by Altschul *et al.* (*Nucl. Acids. Res.* 25: 3389, 1997). A detailed discussion of sequence analysis can be found in Unit 19.3 of Ausubel *et al.* (In: Current Protocols in Molecular Biology, John Wiley & Sons Inc. 1994-1998).

The terms "sequence similarity" and "sequence identity" as used herein refers to the extent that sequences are identical or functionally or structurally similar on a nucleotide-bynucleotide basis or an amino acid-by-amino acid basis over a window of comparison. Thus, a "percentage of sequence identity", for example, is calculated by comparing two optimally aligned sequences over the window of comparison, determining the number of positions at which the identical nucleic acid base (e.g. A, T, C, G, I) or the identical amino acid residue (e.g. Ala, Pro, Ser, Thr, Gly, Val, Leu, Ile, Phe, Tyr, Trp, Lys, Arg, His, Asp, Glu, Asn, Gln, Cys and Met) occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison (i.e., the window size), and multiplying the result by 100 to yield the percentage of sequence identity. For the purposes of the present invention, "sequence identity" will be understood to mean the "match percentage" calculated by the DNASIS computer program (Version 2.5 for windows; available from Hitachi Software engineering Co., Ltd., South San Francisco, California, USA) using standard defaults as used in the reference manual accompanying the software. Similar comments apply in relation to sequence similarity.

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The present invention provides, therefore, an isolated nucleic acid molecule comprising a

sequence of nucleotides selected from SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 and SEQ ID NO:15 or a complementary form thereof. Such nucleic acid molecules encode mammalian homologs of *Drosophila grh*. These mammalian homologs are proposed herein to be transcription factors.

The present invention extends to variants of the nucleic acid molecules. A variant is a molecule having less than 100% sequence identity to a M-grh. Generally, a variant will still hybridize to a M-grh sequence under low stringency conditions.

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The term "variant" refers, therefore, to nucleotide sequences displaying substantial sequence identity with a reference nucleotide sequences or polynucleotides that hybridize with a reference sequence under stringency conditions that are defined hereinafter. The terms "nucleotide sequence", "polynucleotide" and "nucleic acid molecule" may be used herein interchangeably and encompass polynucleotides in which one or more nucleotides have been added or deleted, or replaced with different nucleotides. In this regard, it is well understood in the art that certain alterations inclusive of mutations, additions, deletions and substitutions can be made to a reference nucleotide sequence whereby the altered polynucleotide retains the biological function or activity of the reference polynucleotide.

20 The term "variant" also includes naturally-occurring allelic variants.

Reference herein to a low stringency includes and encompasses from at least about 0 to at least about 15% v/v formamide and from at least about 1 M to at least about 2 M salt for hybridization, and at least about 1 M to at least about 2 M salt for washing conditions.

25 Generally, low stringency is at from about 25-30°C to about 42°C. The temperature may be altered and higher temperatures used to replace formamide and/or to give alternative stringency conditions. Alternative stringency conditions may be applied where necessary, such as medium stringency, which includes and encompasses from at least about 16% v/v to at least about 30% v/v formamide and from at least about 0.5 M to at least about 0.9 M salt for hybridization, and at least about 0.5 M to at least about 0.9 M salt for washing conditions, or high stringency, which includes and encompasses from at least about 31%

v/v to at least about 50% v/v formamide and from at least about 0.01 M to at least about 0.15 M salt for hybridization, and at least about 0.01 M to at least about 0.15 M salt for washing conditions. In general, washing is carried out T<sub>m</sub> = 69.3 + 0.41 (G+C)% (Marmur and Doty, J. Mol. Biol. 5: 109, 1962). However, the T<sub>m</sub> of a duplex DNA decreases by 1°C with every increase of 1% in the number of mismatch base pairs (Bonner and Laskey, Eur. J. Biochem. 46: 83, 1974). Formamide is optional in these hybridization conditions. Accordingly, particularly preferred levels of stringency are defined as follows: low stringency is 6 x SSC buffer, 0.1% w/v SDS at 25°-42°C; a moderate stringency is 2 x SSC buffer, 0.1% w/v SDS at a temperature in the range 20°C to 65°C; high stringency is 0.1 x SSC buffer, 0.1% w/v SDS at a temperature of at least 65°C.

The present invention extends to recombinant forms of the M-grh molecules as well as derivatives and homologs thereof.

Accordingly, another aspect of the present invention provides an isolated polypeptide having transcription factor activity, said polypeptide comprising a sequence of amino acids encoded by a nucleotide sequence selected from SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or a nucleotide sequence having at least about 60% similarity to any one of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or a nucleotide sequence capable of hybridizing to any one of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or a complementary form thereof under low stringency conditions.

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In a preferred embodiment, the present invention provides a recombinant M-grh comprising an amino acid sequence selected from SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14 or SEQ ID NO:16 or an amino acid sequence having at least about 60% similarity to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14 or SEQ ID NO:16.

This aspect of the present invention extends to derivatives, homologs and analogs of M-GRH molecules.

A "derivative" includes a mutant, fragment, part, portion or hybrid molecule. A derivative generally but not exclusively carries a single or multiple amino acid substitution, addition and/or deletion.

A "homolog" includes an analogous polypeptide having at least about 60% similar amino acid sequence from another animal species or from a different locus within the same species.

An "analog" is generally a chemical analog. Chemical analogs of the subject polypeptide contemplated herein include, but are not limited to, modification to side chains, incorporation of unnatural amino acids and/or their derivatives during peptide, polypeptide or protein synthesis and the use of crosslinkers and other methods which impose conformational constraints on the proteinaceous molecule or their analogs.

Examples of side chain modifications contemplated by the present invention include modifications of amino groups such as by reductive alkylation by reaction with an aldehyde followed by reduction with NaBH<sub>4</sub>; amidination with methylacetimidate; acylation with acetic anhydride; carbamoylation of amino groups with cyanate; trinitrobenzylation of amino groups with 2, 4, 6-trinitrobenzene sulphonic acid (TNBS); acylation of amino groups with succinic anhydride and tetrahydrophthalic anhydride; and pyridoxylation of lysine with pyridoxal-5-phosphate followed by reduction with NaBH<sub>4</sub>.

The guanidine group of arginine residues may be modified by the formation of heterocyclic condensation products with reagents such as 2,3-butanedione, phenylglyoxal and glyoxal.

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The carboxyl group may be modified by carbodiimide activation via O-acylisourea formation followed by subsequent derivitization, for example, to a corresponding amide.

Sulphydryl groups may be modified by methods such as carboxymethylation with iodoacetic acid or iodoacetamide; performic acid oxidation to cysteic acid; formation of a mixed disulphides with other thiol compounds; reaction with maleimide, maleic anhydride or other substituted maleimide; formation of mercurial derivatives using 4-chloromercuribenzoate, 4-chloromercuriphenylsulphonic acid, phenylmercury chloride, 2-chloromercuri-4-nitrophenol and other mercurials; carbamoylation with cyanate at alkaline pH.

Tryptophan residues may be modified by, for example, oxidation with N-bromosuccinimide or alkylation of the indole ring with 2-hydroxy-5-nitrobenzyl bromide or sulphenyl halides. Tyrosine residues on the other hand, may be altered by nitration with tetranitromethane to form a 3-nitrotyrosine derivative.

Modification of the imidazole ring of a histidine residue may be accomplished by alkylation with iodoacetic acid derivatives or N-carbethoxylation with diethylpyrocarbonate.

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Examples of incorporating unnatural amino acids and derivatives during peptide synthesis include, but are not limited to, use of norleucine, 4-amino butyric acid, 4-amino-3-hydroxy-5-phenylpentanoic acid, 6-aminohexanoic acid, t-butylglycine, norvaline, phenylglycine, ornithine, sarcosine, 4-amino-3-hydroxy-6-methylheptanoic acid, 2-thienyl alanine and/or D-isomers of amino acids. A list of unnatural amino acid, contemplated herein is shown in Table 3.

# TABLE 3

5	Non-conventional amino acid	Code	Non-conventional amino acid	Code
	α-aminobutyric acid	Abu	L-N-methylalanine	Nmala
	$\alpha$ -amino- $\alpha$ -methylbutyrate	Mgabu	L-N-methylarginine	Nmarg
	aminocyclopropane-	Cpro	L-N-methylasparagine	Nmasn
10	carboxylate		L-N-methylaspartic acid	Nmasp
	aminoisobutyric acid	Aib	L-N-methylcysteine	Nmcys
	aminonorbornyl-	Norb	L-N-methylglutamine	Nmgln
	carboxylate		L-N-methylglutamic acid	Nmglu
	cyclohexylalanine	Chexa	L-Nmethylhistidine	Nmhis
15	cyclopentylalanine	Cpen	L-N-methylisolleucine	Nmile
	D-alanine	Dal	L-N-methylleucine	Nmleu
	D-arginine	Darg	L-N-methyllysine	Nmlys
	D-aspartic acid	Dasp	L-N-methylmethionine	Nmmet
	D-cysteine	Dcys	L-N-methylnorleucine	Nmnle
20	D-glutamine	Dgln	L-N-methylnorvaline	Nmnva
	D-glutamic acid	Dglu	L-N-methylomithine	Nmorn
	D-histidine	Dhis	L-N-methylphenylalanine	Nmphe
	D-isoleucine	Dile	L-N-methylproline	Nmpro
	D-leucine	Dleu	L-N-methylserine	Nmser
25	D-lysine	Dlys	L-N-methylthreonine	Nmthr
	D-methionine	Dmet	L-N-methyltryptophan	Nmtrp
	D-omithine	Dorn	L-N-methyltyrosine	Nmtyr
	D-phenylalanine	Dphe	L-N-methylvaline	Nmval
	D-proline	Dpro	L-N-methylethylglycine	Nmetg
30	D-serine	Dser	L-N-methyl-t-butylglycine	Nmtbug
	D-threonine	Dthr	L-norleucine	Nle

	D-tryptophan	Dtrp	L-norvaline	Nva
	D-tyrosine	Dtyr	α-methyl-aminoisobutyrate	Maib
	D-valine	Dval	α-methyl-γ-aminobutyrate	Mgabu
	D-α-methylalanine	Dmala	α-methylcyclohexylalanine	Mchexa
5	D-α-methylarginine	Dmarg	α-methylcylcopentylalanine	Mcpen
	D-α-methylasparagine	Dmasn	α-methyl-α-napthylalanine	Manap
	D-α-methylaspartate	Dmasp	α-methylpenicillamine	Mpen
	D-α-methylcysteine	Dmcys	N-(4-aminobutyl)glycine	Nglu
	D-α-methylglutamine	Dmgln	N-(2-aminoethyl)glycine	Naeg
10	$D$ - $\alpha$ -methylhistidine	Dmhis	N-(3-aminopropyl)glycine	Norn
	D-α-methylisoleucine	Dmile	N-amino-α-methylbutyrate	Nmaabu
	$D$ - $\alpha$ -methylleucine	Dmleu	$\alpha$ -napthylalanine	Anap
	D- $\alpha$ -methyllysine	Dmlys	N-benzylglycine	Nphe
	$D$ - $\alpha$ -methylmethionine	Dmmet	N-(2-carbamylethyl)glycine	Ngln
15	$D$ - $\alpha$ -methylornithine	Dmorn	N-(carbamylmethyl)glycine	Nasn
	$D$ - $\alpha$ -methylphenylalanine	Dmphe	N-(2-carboxyethyl)glycine	Nglu
	D-α-methylproline	Dmpro	N-(carboxymethyl)glycine	Nasp
	D-α-methylserine	Dmser	N-cyclobutylglycine	Ncbut
-	D-α-methylthreonine	Dmthr	N-cycloheptylglycine	Nchep
20	D-α-methyltryptophan	Dmtrp	N-cyclohexylglycine	Nchex
	$D-\alpha$ -methyltyrosine	Dmty	N-cyclodecylglycine	Ncdec
	D-α-methylvaline	Dmval	N-cylcododecylglycine	Ncdod
	D-N-methylalanine	Dnmala	N-cyclooctylglycine	Ncoct
	D-N-methylarginine	Dnmarg	N-cyclopropylglycine	Ncpro
25	D-N-methylasparagine	Dnmasn	N-cycloundecylglycine	Nound
	D-N-methylaspartate	Dnmasp	N-(2,2-diphenylethyl)glycine	Nbhm
	D-N-methylcysteine	Dnmcys	N-(3,3-diphenylpropyl)glycine	Nbhe
	D-N-methylglutamine	Dnmgln	N-(3-guanidinopropyl)glycine	Narg
	D-N-methylglutamate	Dnmglu	N-(1-hydroxyethyl)glycine	Nthr
30	D-N-methylhistidine	Dnmhis	N-(hydroxyethyl))glycine	Nser

	D-N-methylisoleucine	Dnmile	N-(imidazolylethyl))glycine	Nhis
	D-N-methylleucine	Dnmleu	N-(3-indolylyethyl)glycine	Nhtrp
	D-N-methyllysine	Dnmlys	N-methyl-γ-aminobutyrate	Nmgabu
	N-methylcyclohexylalanine	Nmchexa	D-N-methylmethionine	Dnmmet
5	D-N-methylornithine	Dnmorn	N-methylcyclopentylalanine	Nmcpen
	N-methylglycine	Nala	D-N-methylphenylalanine	Dnmphe
	N-methylaminoisobutyrate	Nmaib	D-N-methylproline	Dnmpro
	N-(1-methylpropyl)glycine	Nile	D-N-methylserine	Dnmser
	N-(2-methylpropyl)glycine	Nleu	D-N-methylthreonine	Dnmthr
10	D-N-methyltryptophan	Dnmtrp	N-(1-methylethyl)glycine	Nval
	D-N-methyltyrosine	Dnmtyr	N-methyla-napthylalanine	Nmanap
	D-N-methylvaline	Dnmval	N-methylpenicillamine	Nmpen
	γ-aminobutyric acid	Gabu	N-(p-hydroxyphenyl)glycine	Nhtyr
	L-t-butylglycine	Tbug	N-(thiomethyl)glycine	Ncys
15	L-ethylglycine	Etg	penicillamine	Pen
	L-homophenylalanine	Hphe	L-α-methylalanine	Mala
	L-α-methylarginine	Marg	L-α-methylasparagine	Masn
	L-α-methylaspartate	Masp	L-α-methyl-t-butylglycine	Mtbug
	L-α-methylcysteine	Mcys	L-methylethylglycine	Metg
20	L-α-methylglutamine	Mgln	$L$ - $\alpha$ -methylglutamate	Mglu
	L-α-methylhistidine	Mhis	$L$ - $\alpha$ -methylhomophenylalanine	Mhphe
	L-α-methylisoleucine	Mile	N-(2-methylthioethyl)glycine	Nmet
	L-α-methylleucine	Mleu	L-α-methyllysine	Mlys
	L-α-methylmethionine	Mmet	L-α-methylnorleucine	Mnle
25	L-α-methylnorvaline	Mnva	L-α-methylornithine	Morn
	L-α-methylphenylalanine	Mphe	L-α-methylproline	Mpro
	L-α-methylserine	Mser	L-a-methylthreonine	Mthr
	L-α-methyltryptophan	Mtrp ·	$L$ - $\alpha$ -methyltyrosine	Mtyr
	L-α-methylvaline	Mval	L-N-methylhomophenylalanine	Nmhpho
3	0 N-(N-(2,2-diphenylethyl)	Nnbhm	N-(N-(3,3-diphenylpropyl)	Nnbhe

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carbamylmethyl)glycine
1-carboxy-1-(2,2-diphenyl- Nmbc
ethylamino)cyclopropane

carbamylmethyl)glycine

Crosslinkers can be used, for example, to stabilize 3D conformations, using homobifunctional crosslinkers such as the bifunctional imido esters having  $(CH_2)_n$  spacer groups with n=1 to n=6, glutaraldehyde, N-hydroxysuccinimide esters and hetero-bifunctional reagents which usually contain an amino-reactive moiety such as N-hydroxysuccinimide and another group specific-reactive moiety such as maleimido or dithio moiety (SH) or carbodiimide (COOH). In addition, peptides can be conformationally constrained by, for example, incorporation of  $C_{\alpha}$  and  $N_{\alpha}$ -methylamino acids, introduction of double bonds

between  $C_{\alpha}$  and  $C_{\beta}$  atoms of amino acids and the formation of cyclic peptides or analogues by introducing covalent bonds such as forming an amide bond between the N and C

15 termini, between two side chains or between a side chain and the N or C terminus.

The present invention further contemplates chemical analogs of the subject polypeptide capable of acting as antagonists or agonists of M-GRH or which can act as functional analogs of M-GRH. Chemical analogs may not necessarily be derived from the instant M-GRH molecules but may share certain conformational similarities. Alternatively, chemical analogs may be specifically designed to mimic certain physiochemical properties of the subject M-GRH molecules. Chemical analogs may be chemically synthesized or may be detected following, for example, natural product screening. The latter refers to molecules identified from various environmental sources such a river beds, coral, plants, microorganisms and insects.

These types of modifications may be important to stabilize the subject M-GRH molecules if administered to an individual or for use as a diagnostic reagent.

30 Other derivatives contemplated by the present invention include a range of glycosylation variants from a completely unglycosylated molecule to a modified glycosylated molecule.

Altered glycosylation patterns may result from expression of recombinant molecules in different host cells.

The present invention also provides a method for identifying a M-GRH, said method comprising screening a nucleotide database and identifying a nucleotide sequence having at least 60% similarity to SEQ ID NO:17 or SEQ ID NO:34 or SEQ ID NO:36 or SEQ ID NO:38 after optimal alignment.

Reference to a "nucleotide database" includes screening an existing genomic or cDNA or mRNA database or screening for a target nucleic acid molecule in a mammalian cell such as using oligonucleotide probes or primers, sequencing the target molecule and comparing the sequence to SEQ ID NO:17 or SEQ ID NO:34 or SEQ ID NO:36 or SEQ ID NO:38.

In an alternative method, a database of mammalian protein sequences is screened for an amino acid sequence having at least 60% similarity to the amino acid sequence encoded by SEQ ID NO:17 or SEQ ID NO:34 or SEQ ID NO:36 or SEQ ID NO:38. Again, a "database" includes a *de novo* protein sequence isolated and identified on a transcription factor isolated form a mammalian cell.

In yet another alternative, a M-grh or its protein product is deemed one which has at least about 60% similarity at the nucleotide level to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or at the amino acid level to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14 or SEQ ID NO:15.

Still yet another aspect of the present invention provides a means of identifying a nucleotide sequence likely to encode an M-GRH transcription factor, said method comprising interrogating a mammalian genome database conceptually translated into different reading frames with an amino acid sequence defining *Drosophila GRH* or any one of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14 and SEQ ID NO:16 and identifying a nucleotide sequence

corresponding to an amino acid sequence having at least about 60% similarity to *Drosophila GRH* or to any one of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14 and SEQ ID NO:16.

Preferably, the genome is conceptually translated into from about 3 to about 6 reading frames and more preferably six reading frames.

It is proposed in accordance with the present invention that the M-GRH transcription factors are involved in the modulation of expression of a number of genes including developmentally regulated genes. Accordingly, aberrations in the M-GRH or M-grh molecules are proposed to cause over or under expression of particular genes leading to a potentially unwanted phenotype. The phenotype may manifest itself pre- or post-natally. A pre-natal manifestation includes at the embryo or fetus stage. Conditions contemplated include developmentally-determined disease conditions such as poor brain development, poor muscle or bone development, aberrations in facial or cranial structures, malformed spinal structures, predispositions to a range of cancers including melanomas and immunological disorders.

Accordingly, another aspect of the present invention contemplates a method for detecting an aberrant phenotype or a propensity for an aberrant phenotype to develop, said method comprising screening for a variation in a nucleotide sequence encoding a mammalian MGR, BOM and/or SOM or their homologs.

Reference herein to "MGR", "BOM" and "SOM" includes murine and human forms of these molecules such as human MGR p49 (SEQ ID NO:2), human MGR p70 (SEQ ID NO:4), human BOM (SEQ ID NO:6), human SOM (SEQ ID NO:8), murine MGR p61 (SEQ ID NO:10), murine MGR p70 (SEQ ID NO:12), murine BOM (SEQ ID NO:14) and murine SOM (SEQ ID NO:16).

A homolog of MGR, BOM and SOM is as herein defined including a molecule having at least about 60% amino acid sequence similarity to MGR, BOM or SOM or at least about

60% nucleic acid similarity to mgr, bom or som or a nucleic acid molecule capable of hybridizing to the coding strands of mgr, bom or som or complementary forms thereof under low stringency conditions.

Aberrations may also be detectable at the amino acid level when the mammalian homologs of *Drosophila grh* encode protein transcription factors.

Accordingly, another aspect of the present invention contemplates a method for detecting an aberrant phenotype or a propensity for an aberrant phenotype to develop, said method comprising screening for a variation in an amino acid sequence encoding MGR, BOM and/or SOM or their homologs.

As above, reference to MGR, BOM and SOM include amino acid sequences defining human MGR p49 (SEQ ID NO:2), human MGR p70 (SEQ ID NO:4), human BOM SEQ ID NO:6), human SOM (SEQ ID NO:8), murine MGR p61 (SEQ ID NO:10), murine MGR p70 (SEQ ID NO:12), murine BOM (SEQ ID NO:14) and murine SOM (SEQ ID NO:16).

As stated above, the mammalian transcription factors and their genetic sequences have a range of diagnostic and therapeutic utilities. The detection of an aberrant transcription factor or a nucleotide sequence encoding an aberrant transcription factor is indicative of a disease condition including a degenerative or developmental disease condition.

Any number of methods may be employed to detect aberrant transcription factors or their genetic sequences. Immunological testing is one particular method. Accordingly, the present invention extends to antibodies and other immunological agents directed to or preferably specific for the mammalian transcription factors or a fragment thereof. The antibodies may be monoclonal or polyclonal or may comprise Fab fragments or synthetic forms.

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Specific antibodies can be used to screen for the subject mammalian transcription factors and/or their fragments. Techniques for the assays contemplated herein are known in the art and include, for example, sandwich assays and ELISA.

It is within the scope of this invention to include any second antibodies (monoclonal, polyclonal or fragments of antibodies or synthetic antibodies) directed to the first mentioned antibodies referred to above. Both the first and second antibodies may be used in detection assays or a first antibody may be used with a commercially available anti-immunoglobulin antibody. An antibody as contemplated herein includes any antibody specific to any region of the mammalian transcription factors.

Both polyclonal and monoclonal antibodies are obtainable by immunization with mammalian transcription factors or antigenic fragments thereof and either type is utilizable for immunoassays. The methods of obtaining both types of sera are well known in the art. Polyclonal sera are less preferred but are relatively easily prepared by injection of a suitable laboratory animal with an effective amount of subject polypeptide, or antigenic parts thereof, collecting serum from the animal and isolating specific sera by any of the known immunoadsorbent techniques. Although antibodies produced by this method are utilizable in virtually any type of immunoassay, they are generally less favoured because of the potential heterogeneity of the product.

The use of monoclonal antibodies in an immunoassay is particularly preferred because of the ability to produce them in large quantities and the homogeneity of the product. The preparation of hybridoma cell lines for monoclonal antibody production derived by fusing an immortal cell line and lymphocytes sensitized against the immunogenic preparation can be done by techniques which are well known to those who are skilled in the art.

Another aspect of the present invention contemplates, therefore, a method for detecting a mammalian transcription factor or fragment thereof in a biological sample from a subject, said method comprising contacting said biological sample with an antibody specific for said mammalian transcription factor or fragment thereof or its derivatives or homologs for

a time and under conditions sufficient for an antibody-polypeptide complex to form, and then detecting said complex.

A biological sample includes a cell extract.

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Reference to a "mammalian transcription factor" is considered to be a reference to a homolog of Drosophia grh, i.e. M-GRH.

The presence of the instant mammalian transcription factors or their fragments may be detected in a number of ways such as by Western blotting and ELISA procedures. A wide range of immunoassay techniques are available as can be seen by reference to U.S. Patent Nos. 4,016,043, 4,424,279 and 4,018,653.

Sandwich assays are among the most useful and commonly used assays and are favoured for use in the present invention. A number of variations of the sandwich assay technique exist, and all are intended to be encompassed by the present invention. Briefly, in a typical forward assay, an unlabeled antibody is immobilized on a solid substrate and the sample to be tested brought into contact with the bound molecule. After a suitable period of incubation, for a period of time sufficient to allow formation of an antibody-antigen 20 complex, a second antibody specific to the antigen, labeled with a reporter molecule capable of producing a detectable signal is then added and incubated, allowing time sufficient for the formation of another complex of antibody-antigen-labeled antibody. Any unreacted material is washed away, and the presence of the antigen is determined by observation of a signal produced by the reporter molecule. The results may either be qualitative, by simple observation of the visible signal, or may be quantitated by comparing with a control sample containing known amounts of hapten. Variations on the forward assay include a simultaneous assay, in which both sample and labeled antibody are added simultaneously to the bound antibody. These techniques are well known to those skilled in the art, including any minor variations as will be readily apparent. In accordance with the present invention the sample is one which might contain a subject transcription factor including by tissue biopsy, blood, synovial fluid and/or lymph. The sample is,

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therefore, generally a biological sample comprising biological fluid. The transcription factor is likely to be in blood or other fluid in the case where cell apoptosis is occurring.

In the typical forward sandwich assay, a first antibody having specificity for the instant polypeptide or antigenic parts thereof, is either covalently or passively bound to a solid surface. The solid surface is typically glass or a polymer, the most commonly used polymers being cellulose, polyacrylamide, nylon, polystyrene, polyvinyl chloride or polypropylene. The solid supports may be in the form of tubes, beads, discs of microplates, or any other surface suitable for conducting an immunoassay. The binding processes are well-known in the art and generally consist of cross-linking covalently binding or physically adsorbing, the polymer-antibody complex is washed in preparation for the test sample. An aliquot of the sample to be tested is then added to the solid phase complex and incubated for a period of time sufficient (e.g. 2-40 minutes or where more convenient, overnight) and under suitable conditions (e.g. for about 20°C to about 40°C) to allow binding of any subunit present in the antibody. Following the incubation period, the antibody subunit solid phase is washed and dried and incubated with a second antibody specific for a portion of the hapten. The second antibody is linked to a reporter molecule which is used to indicate the binding of the second antibody to the hapten.

An alternative method involves immobilizing the target molecules in the biological sample and then exposing the immobilized target to specific antibody which may or may not be labeled with a reporter molecule. Depending on the amount of target and the strength of the reporter molecule signal, a bound target may be detectable by direct labelling with the antibody. Alternatively, a second labeled antibody, specific to the first antibody is exposed to the target-first antibody complex to form a target-first antibody-second antibody tertiary complex. The complex is detected by the signal emitted by the reporter molecule.

By "reporter molecule" as used in the present specification, is meant a molecule which, by its chemical nature, provides an analytically identifiable signal which allows the detection of antigen-bound antibody. Detection may be either qualitative or quantitative. The most commonly used reporter molecules in this type of assay are either enzymes, fluorophores

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or radionuclide containing molecules (i.e. radioisotopes) and chemiluminescent molecules. In the case of an enzyme immunoassay, an enzyme is conjugated to the second antibody, generally by means of glutaraldehyde or periodate. As will be readily recognized, however, a wide variety of different conjugation techniques exist, which are readily available to the skilled artisan. Commonly used enzymes include horseradish peroxidase, glucose oxidase, beta-galactosidase and alkaline phosphatase, amongst others. The substrates to be used with the specific enzymes are generally chosen for the production, upon hydrolysis by the corresponding enzyme, of a detectable colour change. Examples of suitable enzymes include alkaline phosphatase and peroxidase. It is also possible to employ fluorogenic substrates, which yield a fluorescent product rather than the chromogenic substrates noted above. In all cases, the enzyme-labeled antibody is added to the first antibody hapten complex, allowed to bind, and then the excess reagent is washed away. A solution containing the appropriate substrate is then added to the complex of antibody-antigenantibody. The substrate will react with the enzyme linked to the second antibody, giving a qualitative visual signal, which may be further quantitated, usually spectrophotometrically, to give an indication of the amount of hapten which was present in the sample. "Reporter molecule" also extends to use of cell agglutination or inhibition of agglutination such as red blood cells on latex beads, and the like.

Alternately, fluorescent compounds, such as fluorescein and rhodamine, may be 20 chemically coupled to antibodies without altering their binding capacity. When activated by illumination with light of a particular wavelength, the fluorochrome-labeled antibody adsorbs the light energy, inducing a state to excitability in the molecule, followed by emission of the light at a characteristic colour visually detectable with a light microscope. 25 The fluorescent labeled antibody is allowed to bind to the first antibody-hapten complex. After washing off the unbound reagent, the remaining tertiary complex is then exposed to the light of the appropriate wavelength the fluorescence observed indicates the presence of the hapten of interest. Immunofluorescene and EIA techniques are both very well established in the art and are particularly preferred for the present method. However, other reporter molecules, such as radioisotope, chemiluminescent or bioluminescent molecules, 30 may also be employed.

The present invention also contemplates genetic assays such as involving PCR analysis to detect RNA expression products of a genetic sequence encoding a mammalian transcription factor. The genetic assays may also be able to detect nucleotide polymorphisms or other substitutions, additions and/or deletions in the nucleotide sequence of a mammalian transcription factor. Changes in levels of mammalian transcription factor expression such as following mutations in the promoter or regulatory regions or loss of mammalian transcription factor activity following mutations in mammalian transcription factor nucleotides is proposed to be indicative of a disease condition or a propensity for a disease condition to develop. For example, a cell biopsy could be obtained and DNA or RNA extracted. Alternative methods which may be used alone or in conjunction with other methods include direct nucleotide sequencing or mutation scanning such as single stranded conformation polymorphoms analysis (SSCP) as well as specific oligonucleotide hybridization, denaturing high performance liquid chromatography, first nucleotide change 15 (FNC) amongst others.

The present invention extends to polymorphisms which in the M-grh genes leads to healthy or abnormal phenotypes.

The present invention further contemplates kits to facilitate the rapid detection of 20 mammalian transcription factors or their fragments in a subject's biological fluid.

Again, a biological fluid includes a cell extract such as a DNA/RNA extract.

- Still yet another aspect of the present invention contemplates genomic sequences including 25 gene sequences encoding a mammalian transcription factor as well as regulatory regions such as promoters, terminators and transcription/translation enhancer regions associated with the gene encoding a mammalian transcription factor.
- The term "gene" is used in its broadest sense and includes cDNA corresponding to the 30 exons of a gene. Accordingly, reference herein to a "gene" is to be taken to include:-

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- (i) a classical genomic gene consisting of transcriptional and/or translational regulatory sequences and/or a coding region and/or non-translated sequences (i.e. introns, 5'- and 3'- untranslated sequences); or
- (ii) mRNA or cDNA corresponding to the coding regions (i.e. exons) and 5'- and 3'- untranslated sequences of the gene.

The term "gene" is also used to describe synthetic or fusion molecules encoding all or part of an expression product. In particular embodiments, the term "nucleic acid molecule" and "gene" may be used interchangeably.

In a particularly useful embodiment, the present invention provides a promoter for the mammalian transcription factor gene. The identification of the promoter permits developmentally-regulated expression of particular genetic sequences. The latter would include a range of therapeutic molecules such as cytokines, growth factors, antibiotics or other molecules to assist in the treatment of particular disease conditions.

Accordingly, another aspect of the present invention provides a M-grh-specific promoter or functional derivative or homolog thereof, said promoter in situ operably linked to a nucleotide sequence comprising any one of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or their complementary forms or a nucleotide sequence having at least about 60% similarity to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or their complementary forms or a nucleotide sequence capable of hybridizing to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or their complementary forms under low stringency conditions.

30 The promoter is conveniently resident in a vector which comprises unique restriction sites to facilitate the introduction of genetic sequences operably linked to the promoter.

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All such constructs are useful in order to produce recombinant M-GRH molecules and/or in gene therapy protocols.

5 The present invention further contemplates a genetically modified animal.

More particularly, the present invention provides an animal model useful for screening for agents capable of ameliorating the effects of an aberrant M-GRH or M-grh gene. In one embodiment, the animal model produces low amounts of M-grh. Such an animal would have a predisposition for a range of diseases including developmentally regulated diseases. The animal model is useful for screening for agents which ameliorate such conditions.

Accordingly, another aspect of the present invention provides a genetically modified animal wherein said animal produces low amounts of M-grh relative to a non-genetically modified animal of the same species. Reference to "low amounts" includes zero amounts or up to about 10% lower than normalized amounts.

Preferably, the genetically modified animal is a mouse, rat, guinea pig, rabbit, pig, sheep or goat. More preferably, the genetically modified animal is a mouse or rat. Most preferably, the genetically modified animal is a mouse.

Accordingly, a preferred aspect of the present invention provides a genetically modified mouse wherein said mouse produces low amounts of M-grh relative to a non-genetically modified mouse of the same strain.

The animal model contemplated by the present invention comprises, therefore, an animal which is substantially incapable of producing a M-grh. Generally, but not exclusively, such an animal is referred to as a homozygous or heterozygous M-grh-knockout animal.

The animal models of the present invention may be in the form of the animals or may be, for example, in the form of embryos for transplantation. The embryos are preferably maintained in a frozen state and may optionally be sold with instructions for use.

The genetically modified animals may also produce larger amounts of M-GRH For example, over expression of normal M-grh or mutant M-grh may produce dominant negative effects and may become useful disease models.

Accordingly, another aspect of the present invention is directed to a genetically modified animal over-expressing genetic sequences encoding M-grh.

A genetically modified animal includes a transgenic animal, or a "knock-out" or "knock-in" animal.

15 Yet another aspect of the present invention provides a targeting vector useful for inactivating a gene encoding M-GRH, said targeting vector comprising two segments of genetic material encoding said M-GRH flanking a positive selectable marker wherein when said targeting vector is transfected into embryonic stem (ES) cells and the marker selected, an ES cell is generated in which the gene encoding said M-GDH is inactivated by 20 homologous recombination.

Preferably, the ES cells are from mice, rats, guinea pigs, pigs, sheep or goats. Most preferably, the ES cells are from mice.

25 Still yet another aspect of the present invention is directed to the use of a targeting vector as defined above in the manufacture of a genetically modified animal substantially incapable of producing M-GRH.

Even still another aspect of the present invention is directed to the use of a targeting vector as defined above in the manufacture of a genetically modified mouse substantially incapable of producing M-GRH.

Preferably, the vector is DNA. A selectable marker in the targeting vector allows for selection of targeted cells that have stably incorporated the targeting DNA. This is especially useful when employing relatively low efficiency transformation techniques such as electroporation, calcium phosphate precipitation and liposome fusion where typically fewer than 1 in 1000 cells will have stably incorporated the exogenous DNA. Using high efficiency methods, such as microinjection into nuclei, typically from 5-25% of the cells will have incorporated the targeting DNA; and it is, therefore, feasible to screen the targeted cells directly without the necessity of first selecting for stable integration of a selectable marker. Either isogenic or non-isogenic DNA may be employed.

Examples of selectable markers include genes conferring resistance to compounds such as antibiotics, genes conferring the ability to grow on selected substrates, genes encoding proteins that produce detectable signals such as luminescence. A wide variety of such markers are known and available, including, for example, antibiotic resistance genes such as the neomycin resistance gene (neo) and the hygromycin resistance gene (hyg). Selectable markers also include genes conferring the ability to grow on certain media substrates such as the tk gene (thymidine kinase) or the hprt gene (hypoxanthine phosphoribosyltransferase) which confer the ability to grow on HAT medium (hypoxanthine, aminopterin and thymidine); and the bacterial gpt gene (guanine/xanthine phosphoribosyltransferase) which allows growth on MAX medium (mycophenolic acid, adenine and xanthine). Other selectable markers for use in mammalian cells and plasmids carrying a variety of selectable markers are described in Sambrook et al., Molecular Cloning - A Laboratory Manual, Cold Spring Harbour, New York, USA, 1990.

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The preferred location of the marker gene in the targeting construct will depend on the aim of the gene targeting. For example, if the aim is to disrupt target gene expression, then the selectable marker can be cloned into targeting DNA corresponding to coding sequence in the target DNA. Alternatively, if the aim is to express an altered product from the target gene, such as a protein with an amino acid substitution, then the coding sequence can be

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modified to code for the substitution, and the selectable marker can be placed outside of the coding region, for example, in a nearby intron.

The selectable marker may depend on its own promoter for expression and the marker gene may be derived from a very different organism than the organism being targeted (e.g. prokaryotic marker genes used in targeting mammalian cells). However, it is preferable to replace the original promoter with transcriptional machinery known to function in the recipient cells. A large number of transcriptional initiation regions are available for such purposes including, for example, metallothionein promoters, thymidine kinase promoters,  $\beta$ -actin promoters, immunoglobulin promoters, SV40 promoters and human cytomegalovirus promoters. A widely used example is the pSV2-neo plasmid which has the bacterial neomycin phosphotransferase gene under control of the SV40 early promoter and confers in mammalian cells resistance to G418 (an antibiotic related to neomycin). A number of other variations may be employed to enhance expression of the selectable markers in animal cells, such as the addition of a poly(A) sequence and the addition of synthetic translation initiation sequences. Both constitutive and inducible promoters may be used.

The DNA is preferably modified by homologous recombination. The target DNA can be in any organelle of the animal cell including the nucleus and mitochondria and can be an intact gene, an exon or intron, a regulatory sequence or any region between genes.

Homologous DNA is a DNA sequence that is at least 70% identical with a reference DNA sequence. An indication that two sequences are homologous is that they will hybridize with each other under stringent conditions (Sambrook et al., 1990, supra).

The present invention further contemplates co-suppression (i.e. sense suppression) and antisense suppression to down-regulate expression of M-grh This would generally occur in a target test animal such as to generate a disease model.

In addition to providing a diagnostic capability as described above, the isolated nucleic

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acid molecules of the present invention may also provide a therapeutic capability by being used to correct or complement an abnormality detected in a subject. To deliver the appropriate sequence to a recipient cell or tissue of a subject, an isolated nucleic acid molecule of the present invention may be cloned into a suitable genetic construct such as a suitable vector.

Accordingly, a further aspect of the present invention contemplates a genetic construct comprising a nucleotide sequence encoding an M-grh selected from SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or a variant thereof or a nucleotide sequence having at least 60% similarity to one or more of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13 or SEQ ID NO:15 or a variant thereof or a nucleotide sequence capable of hybridizing to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:15 under low stringency conditions or a variant thereof or a complementary form thereof.

A "vector" is a polynucleotide molecule, preferably a DNA molecule derived, for example, from a plasmid, bacteriophage, or plant virus, into which a polynucleotide can be inserted or cloned. A vector preferably contains one or more unique restriction sites and can be capable of autonomous replication in a defined host cell including a target cell or tissue or a progenitor cell or tissue thereof, or be integrable with the genome of the defined host such that the cloned sequence is reproducible. Accordingly, the vector may be an autonomously replicating vector, i.e. a vector that exists as an extra-chromosomal entity, the replication of which is independent of chromosomal replication. Examples include a linear or closed circular plasmid, an extra-chromosomal element, a mini-chromosome, or an artificial chromosome. The vector may also contain a means for assuring self-replication. Alternatively, the vector may be one which, when introduced into the host cell, is integrated into the genome and replicated together with the chromosome(s) into which it has been integrated. A vector system may comprise a single vector or plasmid, two or more vectors or plasmids, which together contain the total DNA to be introduced into the genome of the host cell, or a transposon.

Vectors suitable for gene therapy applications are well known in the art. The choice of the vector will typically depend on the compatibility of the vector with the host cell into which it is to be introduced. The vector may also include an additional genetic construct comprising a selection marker such as an antibiotic resistance gene that can be used for selection of suitable transformants. Examples of such resistance genes are known to those skilled in the art and include the *nptII* gene that confers resistance to the antibiotics kanamycin, and G418 (Geneticin®) and the *hph* gene which confer resistance to the antibiotic hygromycin B.

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Accordingly, in a related embodiment, the present invention provides a genetic construct comprising a promoter or functional equivalent thereof operably linked to a nucleotide sequence of the invention.

Reference herein to a "promoter" is to be taken in its broadest context and includes the transcriptional regulatory sequences of a classical genomic gene, which is required for accurate transcription initiation, with or without a CCAAT box sequence and additional regulatory elements (i.e. upstream activating sequences, enhancers and silencers), which alter gene expression in response to developmental and/or external stimuli, or in a tissue-specific manner. A promoter is usually, but not necessarily, positioned upstream (5') of a gene region, the expression of which it regulates. Furthermore, the regulatory elements comprising a promoter are usually positioned within 2 kb of the start site of transcription of the gene. As is known in the art, some variation in this distance can be accommodated without loss of promoter function.

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The selection of an appropriate promoter sequence to regulate expression of a transcription factor encoded by an isolated nucleic acid molecule of the present invention is an important consideration. Examples of suitable promoters include viral, fungal, bacterial, animal and plant derived promoters capable of functioning in eukaryotic animal cells and, especially, human cells. The promoter may regulate the expression of the nucleic acid molecule differentially with respect to the cell, tissue or organ in which expression occurs,

or with respect to the developmental stage at which expression occurs.

Preferably, the promoter is capable of regulating expression of a nucleic acid molecule in a eukaryotic cell, tissue or organ, at least during the period of time over which the regulated gene is expressed therein, and more preferably also immediately preceding the commencement of detectable expression of the regulated gene in said cell, tissue or organ.

Particularly preferred promoters for use with the nucleic acid molecules of the present invention include the bacteriophage T7 promoter, bacteriophage T3 promoter, SP6 promoter, *lac* operator-promoter, *tac* promoter, SV40 late promoter, SV40 early promoter, RSV-LTR promoter, CMV IE promoter, CaMV 35S promoter, SCSV promoter, SCBV promoter and the like. Those skilled in the art will readily be aware of additional promoter sequences other than those specifically described.

- In the present context, the terms "in operable connection with" or "operably linked" or similar shall be taken to indicate that expression of the nucleic acid molecule is under the control of the promoter sequence, with which it is spatially connected, in a cell, tissue, organ or whole organism.
- The genetic construct of the present invention may also comprise a 3' non-translated sequence. A 3' non-translated sequence refers to that portion of a gene comprising a DNA segment that contains a polyadenylation signal and any other regulatory signals capable of effecting mRNA processing or gene expression. The polyadenylation signal is characterized by effecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor. Polyadenylation signals are commonly recognized by the presence of homology to the canonical form 5' AATAAA-3' although variations are not uncommon.

Accordingly, a genetic construct comprising a nucleic acid molecule of the present invention, operably linked to a promoter, may be cloned into a suitable vector for delivery to a cell or tissue in which regulation is faulty, malfunctioning or non-existent, in order to rectify and/or provide the appropriate regulation. Vectors comprising appropriate genetic

constructs may be delivered into target eukaryotic cells by a number of different means well known to those skilled in the art of molecular biology.

The present invention further contemplates the use of an M-GRH or M-grh in the manufacture of a medicament for the treatment of a disease condition in a mammal such as a human.

The present invention is further directed to promoters and 3'- and 5'-regulatory regions associated with genomic forms of M-grh genes. These regions can be readily identified by, for example, chromosome walking using M-grh nucleic acid molecules or probes or primers therefrom.

The present invention is further described by the following non-limiting Examples.

#### **EXAMPLE 1**

#### Polymerase chain reaction

For RT-PCR, first strand cDNA was prepared from 2 µg of mRNA from primary tissues using random hexamers. Each cDNA sample was appropriately diluted to give similar amplification of S14 RNA under the same PCR conditions. The primer sequences are detailed below. The PCR conditions were 94°C for 2 min followed by 35 cycles of 94°C for 30 sec, 60°C for 30 sec and 72°C for 45 sec with a final extension at 72°C for 5 min. All PCR products were electrophoresed on 1.5% w/v agarose gels, transferred to 10 nitrocellulose and analyzed by Southern blot using <sup>32</sup>P-radiolabeled internal oligonucleotides as probes. Membranes were then autoradiographed for 2 hr at -70°C.

The following primers were used to amplify probes for cDNA library screening and for RT-PCR:-

[SEQ ID NO:25]

[SEQ ID NO:26]

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	human p49 mgr	•
	5'-GAAGTCTTTGATGCCCTGATG-3'	[SEQ ID NO:19]
	5'-AACCCATTCCCTCGACATAGA-3'	[SEQ ID NO:20]
20	human p70 mgr	
	5-AGCGCGATGACACAGGAGTA-3'	[SEQ ID NO:21]
	5'-CGTTGCTATGGAGACAGTGA-3'	[SEQ ID NO:22]
	human bom	
25	5'-CCGTTTAACAAGGACACTGC-3'	[SEQ ID NO:23]
	5'-CTGGAAGCCACCAAATCTCT-3'	[SEQ ID NO:24]
	murine p70 mgr	

5'-AGCGCGATGACACAGGAGTA-3'

5'-AGTGCCAGAGCTGAACTGAT-3'

#### murine p61 mgr

5'-TCCATGGGTTCCTTGAGTTC-3' [SEQ ID NO:27] 5'-AGTGCCAGAGCTGAACTGAT'-3' [SEQ ID NO:28]

#### 5 murine bom

5'-AAAGGGGAGCGAGTTCATTG-3' [SEQ ID NO:29] 5'-AGAGCTCTCGGTGATGGATA-3' [SEQ ID NO:30]

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#### **EXAMPLE 2**

### Cloning of human and murine mgr and bom

Human p49 mgr was cloned from a fetal brain cDNA phage library in the λZAP II vector (Stratagene). The cDNA encoding the longer human MGR isoform was amplified by RT-PCR from human kidney mRNA. The cDNA encoding the smaller murine isoform of MGR was cloned from a 17.5-day embryo phage library in the Lambda TripelEx vector (Clontech). The murine p70 cDNA was amplified from murine kidney mRNA by RT-PCR. The human bom cDNA was isolated form a placental phage library in the Lambda ZAP II vector (Stratagene) and the murine cDNA from an embryonic carcinoma cell line (P19) phage library in the Uni-ZAP XR vector (Stratagene). The murine MGR genomic locus was obtained from a 129SVJ phage library in the Lambda FIX II vector (Stratagene).

From similarity searches of GenBank databases, using the GRH protein sequence as a query, two murine expressed sequence tag (EST) entries were found from adult brain and ovary and one human EST entry from fetal brain that were not identical to any previously reported genes, yet shared high degrees of homology with each other and grh. These sequences were used to design murine and human primers and amplified probes from murine adult brain and ovary and human adult brain cDNA. The murine probe from adult brain cDNA was used in a screen of a day 17.5 mouse embryo cDNA library to obtain a full length clone of a gene referred to as mammalian grainyhead (mgr) due to its sequence and functional homology and similar expression pattern to that of the fly gene. The human

probe derived from adult brain cDNA was used to obtain a full length cDNA clone from a human fetal brain library. Amino acid sequence comparison reveals this to be the human homolog of MGR with 94% identity at the amino acid level.

- The murine probe derived from ovary cDNA was used in a screen of a murine teratocarcinoma cell line (P19) cDNA library to obtain a full length clone of a novel gene distinct from but highly related to mgr named brother-of-mgr (bom). The homology between mgr and bom suggests that mgr and bom arose through gene duplication.
- The human homolog of bom was obtained using primers derived from a high throughput 10 genome sequencing (HTGS) database entry with homology to murine bom. These were used to amplify a probe from a human placental cDNA library that was then screened to yield a full length human cDNA clone. Amino acid sequence comparison between murine and human BOM revealed 94% identity.

The sequence alignments between grh, mgr, bom, CP2 and LBP-1a revealed that mgr and bom are more closely related to grh than the previously identified homologs CP2 and LBP-. 1a (Table 4). This homology is particularly evident in the DNA binding and dimerization domains emphasizing the importance of protein/protein and protein/DNA interactions for the function of these factors.

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TABLE 4 Amino acid sequence comparison of GRH-like genes and Drosophila grh

Amino acid identity/ similarity to Grainyhead (%)	Overall	DNA-binding domain	Dimerization domain
MGR	37/52	48/64	39/61
BOM	35/52	46/63	37/61
SOM	33/48	42/60	38/57
CP2	26/42	32/52	29/47
LBP-1a	23/39	31/51	28/43

# EXAMPLE 3 Identification of a second isoform of MGR

A striking feature of the alignment between MGR and BOM was the absence of an MGR domain corresponding to the first 93 amino acids of BOM. In view of the absence of tissue-specific isoforms of GRH, the EST database was searched for similar sequences using the 5' end of bom as a query. A highly similar but non-identical sequence in an EST from murine kidney was located. The most 3' 30 nucleotides of this EST was identical to 30 nucleotides close to the 5' end of the mgr. Based on this, primers were designed from the kidney EST and mgr cDNA sequences and amplified a product of the predicted size from murine kidney cDNA. A similar product was also amplified from human kidney cDNA. Amino acid sequence analysis of the murine product revealed that it was highly homologous to the 5' end of the BOM protein and contiguous with the mgr open reading frame. However, it lacked the first 11 amino acids of a previously isolated mgr clone suggesting the presence of alternate splicing. To examine this, the murine mgr genomic locus was isolated and mapped. As shown in Figure 1B, the first three coding exons in the locus are exclusive to the p70 isoform of mgr. In contrast, the shorter isoform of mgr's (p61) first coding exon is absent in the p70 isoform. Significantly, the 5' end of this exon lacks a splice acceptor site explaining its absence from the longer isoform. Instead, promoter sequences with a clear TATA box and CAP site are evident in close proximity to the translation initiation site (Figure 1C). Subsequent mapping of the human genomic locus revealed that murine exon four was conserved in the human p70 protein but was absent in

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the 49 kDa isoform of MGR.

#### **EXAMPLE 4**

## The first three exons of the mgr genomic locus encode transcriptional activation domain

Although significant sequence homology exists between grh and the shorter mgr isoforms and p70 mgr, the isoleucine rich transcriptional activation domain identified in the fly protein is not conserved. Examination of the MGR-coding sequences failed to reveal a region homologous to other known transactivation domains. In view of the high degree of conservation of the first three coding exons of p70 mgr and bom, it was postulated that this could be the functional domain responsible for activation. To address this, the cDNA fragment encoding the first 93 amino acids of human p70 MGR (encoded by the first three exons) was subcloned in frame with the GAL4 DNA binding domain in a mammalian expression vector. The comparable region of BOM and the full length p49 MGR cDNA in frame into this vector was also cloned. These plasmids were co-transfected into the human 293T cell line with a reporter plasmid containing five concatamerized GAL4 DNA binding sites upstream of the chloramphenicol acetyltransferase (CAT) gene. The vector containing only the GAL4 DNA-BD or containing the VP16 activation domain fused to the GAL DNA-BD served as the negative and positive controls, respectively. As shown in Figure 3, transcriptional activation of the CAT gene was observed with VP16, p70 MGR and the bom containing plasmids. No activation was observed with p49 mgr or the empty vector. These findings confirm the presence of a highly conserved activation domain in the p70 mgr and bom that is lacking in p49 mgr.

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#### **EXAMPLE 5**

#### MGR binds to known GRH binding sites

To determine the extent of the functional homology between GRH and MGR, it was initially examined whether the mammalian protein could bind to the well-characterized binding sites for the *Drosophila* factor in the *Dopa decarboxylase* and *PCNA* gene

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regulatory regions (Uv et al., Mol. Cell. Biol. 17: 6727-6735, 1997; Hayashi et al., J. Biol. Chem. 274: 35080-35088, 1999). Oligonucleotide probes encompassing these sites were incubated with nuclear extract from the human placental cell line JEG-3, which expresses both isoforms of MGR at RNA and protein level and analyzed in an electrophoretic mobility shift assay (EMSA).

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As shown in Figure 2A, a specific protein/DNA complex was observed with the *PCNA* probe in the presence of pre-immune sera (lanes 1 and 3). This complex was supershifted with the addition of anti-p70 specific antisera raised against peptides in the amino terminal region of the protein (lane 4) and ablated with the addition of anti-MGR antisera raised against peptides common to p49 and p70 MGR in the dimerization domain of the protein (lane 2). Neither antisera cross-reacted with BOM. Similar results were obtained with the *Dopa decarboxylase* promoter probe (Figure 2B).

#### **EXAMPLE 6**

#### MGR binds to the human Engrailed-1 promoter

Many Drosophila genes regulated by GRH have known mammalian homologs. In terms of functional homology, Engrailed-1 (En-1) is one of the bests characterized. The En-1 promoter was, therefore, examined for the grainyhead consensus DNA binding sequence derived from a comparison of the Drosophila Ultrabithorax, Dopa decarboxylase and fushi tarazu promoters (Dynlacht et al., Genes Dev. 3: 1677-1688, 1989). As shown in Figure 3A, a highly conserved region was identified in the proximal En-1 promoter. Moreover, this sequence was also largely conserved in the DNAseI footprint attributed to grh in the Drosophila engrailed promoter (Soeller et al., Genes Dev. 2: 68-81, 1988). The ability of this region of the human En-1 promoter to compete off MGR binding to the Ddc probe (Figure 3B) in an EMSA with nuclear extract from JEG-3 cells was examined. As shown in Figure 3B, the specific MGR/DNA complex observed with the Ddc probe (lane 15 1) was supershifted with the addition of MGR antisera 67 (lane 2) and ablated with the addition of a 50-fold excess of unlabeled Ddc probe as competitor (lane 3). Addition of a 10- (lane 4) or 20-fold (lane 5) excess of unlabeled En-1 probe also markedly reduced the binding of MGR to the Ddc probe.

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#### **EXAMPLE 7**

#### MGR activates transcription

To determine the functional significance of this binding, this region of the En-1 promoter was linked to a minimal globin gene promoter/luciferase reporter gene construct and transfected it into the MGR null cell line COS, in the presence of p70 MGR mammalian expression vector or the empty vector. Transfection of the minimal promoter/reporter or the TK promoter linked to a Renilla luciferase gene with either vector served as the controls. As shown in Figure 3C, expression of p70 MGR dramatically enhanced the transcriptional activity of the En-1 promoter (solid bars) but not the control minimal promoter (open bars) or the TK promoter (hatched bars).

#### **EXAMPLE 8**

#### Cloning of full-length human SOM

Human SOM was cloned using primers derived from a high through-put genomic sequence (HTGS) and a human expression sequence tag (EST) obtained from GenBank databases which, respectively, aligned with the dimerization domain and the activation domain of other MGR members. Using nested RT-PCR and human tonsil cDNA, another contig spanning 1300 nucleotides was obtained.

10 Utilizing 5' RACE, further oligoprimers and human testis cDNA, a 210 nucleotide sequence incorporating the initiating ATG was obtained. A contig of these overlapping sequences revealed the full length human SOM which upon alignment with other existing MGR family members showed >60% similarity at the protein level with conservation at the 5' activation, DNA-binding and dimerization domains.

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#### **EXAMPLE 9**

#### Cloning of full-length murine SOM

A murine EST (GenBank) from optic cup tissue was identified, which when aligned with other murine homologs of the MGR family showed 70% similarity at the amino acid level, in the region of the DNA binding domain. Using semi-nested RT-PCR with murine testis cDNA, a 286 nucleotide sequence was amplified, cloned and sequenced for use as a probe.

Subsequently, a murine brain cDNA library (Stratagene) was screened. One clone was taken through to quaternary stage. This clone was excised from λZAP II vector into pBluescript and sequenced in both directions. A 1200 nucleotide length sequence was obtained, whichi lacked the 5' end. This was subsequently identified using 5' RACE from murine testis cDNA. A contig of these two sequences revealed the full length murine SOM.

Those skilled in the art will appreciate that the invention described herein is susceptible to variations and modifications other than those specifically described. It is to be understood that the invention includes all such variations and modifications. The invention also includes all of the steps, features, compositions and compounds referred to or indicated in this specification, individually or collectively, and any and all combinations of any two or more of said steps or features.

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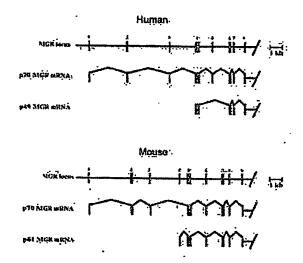
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### Figure 1A



## Figure 1B

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Figure 1C

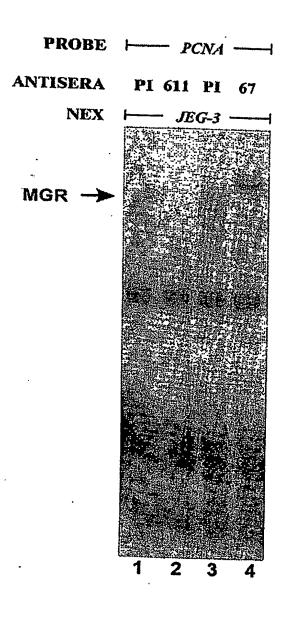


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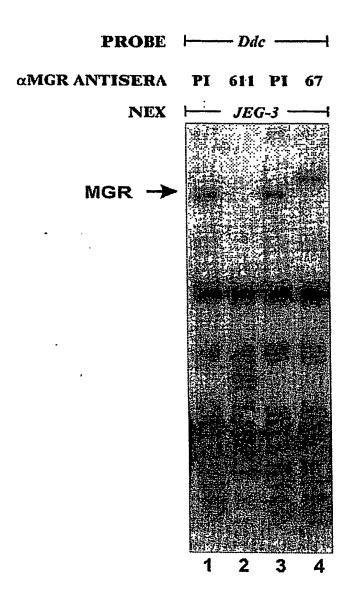


Figure 2B

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## Figure 3A

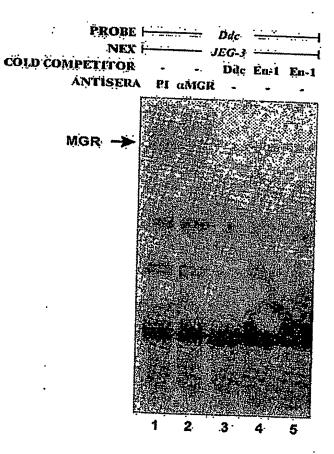


Figure 3B

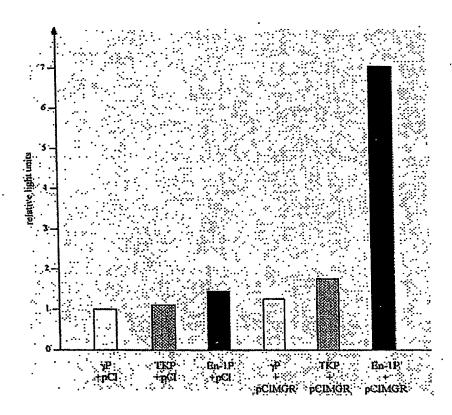


Figure 3C

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gtg aag cat tac tcc aat gag gac acc ttc cag ctg cag att gaa gaa Val Lys His Tyr Ser Asn Glu Asp Thr Phe Gln Leu Gln Ile Glu Glu 595 600 605	1824
gcc ggg ggg tct tac aag ctc acc ctg acg gag atc taaaggcctg Ala Gly Gly Ser Tyr Lys Leu Thr Leu Thr Glu Ile 610 615	1870
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Met Ser Ile Asn Gly Asp Glu Asp Ser Ala Ala Ala Leu Gly Leu Leu 50 55 60	
Tyr Asp Tyr Tyr Lys Val Pro Arg Glu Arg Arg Ser Ser Thr Ala Lys 65 70 75 80	

Pro Glu Val Glu His Pro Glu Pro Asp His Ser Lys Arg Asn Ser Ile

Pro Ile Val Thr Glu Gln Pro Leu Ile Ser Ala Gly Glu Asn Arg Val-105 Gln Val Leu Lys Asn Val Pro Phe Asn Ile Val Leu Pro His Gly Asn Gln Leu Gly Ile Asp Lys Arg Gly His Leu Thr Ala Ser Asp Thr Thr Val Thr Val Ser Ile Ala Thr Met Pro Thr His Ser Ile Lys Thr Glu Thr Gln Pro His Gly Phe Ala Val Gly Ile Pro Pro Ala Val Tyr His Pro Glu Pro Thr Glu Arg Val Val Val Phe Asp Arg Asn Leu Asn Thr 180 Asp Gln Phe Ser Ser Gly Ala Gln Ala Pro Asn Ala Gln Arg Arg Thr 200 Pro Asp Ser Thr Phe Ser Glu Thr Phe Lys Glu Gly Val Gln Glu Val Phe Phe Pro Ser Asp Leu Ser Leu Arg Met Pro Gly Met Asn Ser Glu 230 225 . Asp Tyr Val Phe Asp Ser Val Ser Gly Asn Asn Phe Glu Tyr Thr Leu 250 Glu Ala Ser Lys Ser Leu Arg Gln Lys Pro Gly Asp Ser Thr Met Thr Tyr Leu Asn Lys Gly Gln Phe Tyr Pro Ile Thr Leu Lys Glu Val Ser Ser Ser Glu Gly Ile His His Pro Ile Ser Lys Val Arg Ser Val Ile Met Val Val Phe Ala Glu Asp Lys Ser Arg Glu Asp Gln Leu Arg His

Ala Lys Val Phe Ile Ser Val Asn Cys Leu Ser Thr Asp Phe Ser Ser

Glu Ile Ala Tyr Asn Ala Ile Ser Phe Thr Trp Asp Ile Asn Asp Glu 355 360 365

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CDS

(67) . . (1941)

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gtg ccc atg ccc agt gac cct cca ttc aat acc cga aga gcc tac acc Val Pro Met Pro Ser Asp Pro Pro Phe Asn Thr Arg Arg Ala Tyr Thr 15 20 25 30	<b>156</b>									
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gca Ala	gcc Ala	aca Thr 225	gag Glu	aaa Lys	ttt Phe	cgg Arg	agt Ser 230	gct Ala	tca Ser	gtt Val	gjå aaa	gct Ala 235	gag Glu	gag Glu	tac Tyr	780
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Tyr	320	His	Ser	Arg	Gln	His 325	Thr	gcg Ala	Lys	Gln	Arg 330	Val	Leu	Asp	Ile	1068
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	20			25 ′			;	30			
Asp Glu Al	a Trp L	ys Ser	Tyr Leu 40	Glu Asn	Pro		Thr i	Ala	Ala	Thr	
Lys Ala Me 50	t Met S	er Ile	Asn Gly 55	Asp Glu	Asp	Ser 60	Ala 2	Ala	Ala	Leu	-
Gly Len Le											
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65 Ser Val Se	r Lys A	70			75					80	
05	r Lys A 8	70 la Ser 5	Asp Ser	Gln Glu 90	75 Asp	Gln	Glu :	Lys	Arg 95	80 Asn	

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Tyr	Tyr	Asn	Thr 500	Asp	Asp	. Glu	Arg	Glu 505	Gly	Gly	Ser	Val	Leu 510	Val	Lys
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Gln	Met 530	ГÀв	Glu	Glu	Gly	Thr 535	Lys	Arg	Val	Leu	Leu 540	Tyr	Val	Arg	Lys
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ГЛе	Gly	Leu	Met	Glu 565		Ile	Ser	Glu	Ьуя 570	_	Gly	Leu	Pro	Val 575	Glu
Lys	Ile	Ala	Lys 580		Tyr	ГЛЯ	Lys	Ser 585		Lys	Gly	Ile	Leu 590		Asn
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221	+ +c	r at	t ct	+ cc			+ <+	<b>-</b>	~ ~~	~ +~	- ~-	~ ~~	~ ~+	~ ^+	

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Gln Lys Gly Val Lys Gly Leu Pro Leu Asn Ile Gln Ile Asp Thr Tyr 385 390 395 400

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Ile Lys Val Phe Cys Asp Lys Gly Ala Glu Arg Lys Ile Arg Asp Glu

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Pro Asp Ile Leu Lys Thr Ser Pro Asp Pro Pro Cys Pro Glu Asp Tyr 210 215 220

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Ile His Ile Lys Ala Gly Glu Ser Pro Met Ala Tyr Leu Asn Lys Gly

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Val Met Pro Ala Asp Gly Gly Gly Gly Asn Asn Ser Asp Glu Gly His 130 135 140

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Gln Gln Gln Gln Gln Gln Thr Glu His Gln Pro Leu Ala Lys Ile 165 170 175

Glu Phe Asp Glu Asn Gln Ile Ile Arg Val Val Gly Pro Asn Gly Glu 180 185 190

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Gly Ile Gly Ala Ala Thr Gly Met Thr Phe Asn Pro Leu Ser Asn Gly 865 870 875 880

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Gly Gln Lys Val Ala Thr Pro Thr Leu Lys Phe His Asn His Phe Pro 915 920 925

Pro Asp Met Gln Thr Asp Lys Lys Asp His Ile Leu Asp Gln Asn Met 930 935 940

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His Lys Lys His Ala Ala Ala Ala Ala Ala Ala Gly Gly Gly Ser

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His Asp Phe Gly Arg Gln Asn Asp Ala Asp Lys Ile Gln Ile Pro Lys 885 Ile Phe Thr Asn Val Gly Phe Arg Tyr His Leu Glu Ser Pro Ile Ser Ser Ser Gln Arg Arg Glu Asp Asp Arg Ile Thr Tyr Ile Asn Lys Gly Gln Phe Tyr Gly Ile Thr Leu Glu Tyr Val His Asp Ala Glu Lys Pro Ile Lys Asn Thr Thr Val Lys Ser Val Ile Met Leu Met Phe Arg Glu Glu Lys Ser Pro Glu Asp Glu Ile Lys Ala Trp Gln Phe Trp His Ser 965 Arg Gln His Ser Val Lys Gln Arg Ile Leu Asp Ala Asp Thr Lys Asn Ser Val Gly Leu Val Gly Cys Ile Glu Glu Val Ser His Asn Ala Ile 995 Ala Val Tyr Trp Asn Pro Leu Glu Ser Ser Ala Lys Ile Asn Ile 1010 1015 1020 Ala Val Gln Cys Leu Ser Thr Asp Phe Ser Ser Gln Lys Gly Gly 1030 Leu Pro Leu His Val Gln Ile Asp Thr Phe Glu Asp Pro Arg Asp Thr Ala Val Phe His Arg Gly Tyr Cys Gln Ile Lys Val Phe Cys 1060 1065 Asp Lys Gly Ala Glu Arg Lys Thr Arg Asp Glu Glu Arg Arg Ala 1070 Ala Lys Arg Lys Met Thr Ala Thr Gly Arg Lys Lys Leu Asp Glu Leu Tyr His Pro Val Thr Asp Arg Ser Glu Phe Tyr Gly Met Gln 1100 1105 1110 Asp Phe Ala Lys Pro Pro Val Leu Phe Ser Pro Ala Glu Asp Met 1120 Glu Lys Val Gly Gln Leu Gly Ile Gly Ala Ala Thr Gly Met Thr 1135 Phe Asn Pro Leu Ser Asn Gly Asn Ser Asn Ser Asn Ser His Ser 1145 1150

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250 Gln Thr Ser Pro Leu Pro Leu Asp Ala Ser Val Leu His Tyr Ser Gly Gly Asn Asp Ser Asn Val Ile Lys Thr Glu Ala Asp Ile Tyr Glu Asp 280 His Lys Lys His Ala Ala Ala Ala Ala Ala Ala Gly Gly Gly Ser Ile Ile Tyr Thr Thr Ser Asp Pro Asn Gly Val Asn Val Lys Gln Leu 310 Pro His Leu Thr Val Pro Gln Lys Leu Asp Pro Asp Leu Tyr Gln Ala 330 Asp Lys His Ile Asp Leu Ile Tyr Asn Asp Gly Ser Lys Thr Val Ile Tyr Ser Thr Thr Asp Gln Lys Ser Leu Glu Ile Tyr Ser Gly Gly Asp 355 Ile Gly Ser Leu Val Ser Asp Gly Gln Val Val Val Gln Ala Gly Leu Pro Tyr Ala Thr Thr Gly Ala Gly Gly Gln Pro Val Tyr Ile Val Ala Asp Gly Ala Leu Pro Ala Gly Val Glu Glu His Leu Gln Ser Gly 410 405 Lys Leu Asn Gly Gln Thr Thr Pro Ile Asp Val Ser Gly Leu Ser Gln Asn Glu Ile Gln Gly Phe Leu Leu Gly Ser His Pro Ser Ser Ser Ala Thr Val Ser Thr Thr Gly Val Val Ser Thr Thr Thr Ile Ser His His Gln His Gln Gln Gln Gln His Pro Gly Asp Ile Val Ser Ala Ala 490 Gly Val Gly Ser Thr Gly Ser Ile Val Ser Ser Ala Ala Gln Gln 500 Gln Gln Gln Leu Ile Ser Ile Lys Arg Glu Pro Glu Asp Leu Arg Lys Asp Pro Lys Asn Gly Asn Ile Ala Gly Ala Ala Thr Ala Asn Gly

535 540 530 Pro Gly Ser Val Ile Thr Gln Lys Ile Leu His Val Asp Ala Pro Thr Ala Ser Glu Ala Asp Arg Pro Ser Thr Pro Ser Ser Ile Asn Ser 565 570 Thr Glu Asn Thr Glu Ser Asp Ser Gln Ser Val Ser Gly Ser Glu Ser Gly Ser Pro Gly Ala Arg Thr Thr Ala Thr Leu Glu Met Tyr Ala Thr 600 Thr Gly Gly Thr Gln Ile Tyr Leu Gln Thr Ser His Pro Ser Thr Ala Ser Gly Ala Gly Gly Gly Ala Gly Pro Ala Gly Ala Ala Gly Gly Gly Gly Val Ser Met Gln Ala Gln Ser Pro Ser Pro Gly Pro Tyr Ile Thr 650 Ala Asn Asp Tyr Gly Met Tyr Thr Ala Ser Arg Leu Pro Pro Gly Pro Pro Pro Thr Ser Thr Thr Thr Phe Ile Ala Glu Pro Ser Tyr Tyr Arg Glu Tyr Phe Ala Pro Asp Gly Gln Gly Gly Tyr Val Pro Ala Ser Thr 695 Arg Ser Leu Tyr Gly Asp Val Asp Val Ser Val Ser Gln Pro Gly Gly Val Val Thr Tyr Glu Gly Arg Phe Ala Gly Ser Val Pro Pro Pro Ala Thr Thr Thr Val Leu Thr Ser Val His His His Gln Gln Gln Gln 745 Gln Gln Gln His Gln Gln Gln Gln Gln Gln Gln His His Gln Gln Gln Gln His His Ser Gln Asp Gly Lys Ser Asn Gly Gly Ala Thr Pro Leu Tyr Ala Lys Ala Ile Thr Ala Ala Gly Leu Thr Val Asp Leu Pro Ser Pro Asp Ser Gly Ile Gly Thr Asp Ala Ile Thr Pro Arg Asp 810 Gln Thr Asn Ile Gln Gln Ser Phe Asp Tyr Thr Glu Leu Cys Gln Pro Gly Thr Leu Ile Asp Ala Asn Gly Ser Ile Pro Val Ser Val Asn Ser 835 840 845

Ile Gln Gln Arg Thr Ala Val His Gly Ser Gln Asn Ser Pro Thr Thr 850 855 860

Ser Leu Val Asp Thr Ser Thr Asn Gly Ser Thr Arg Ser Arg Pro Trp 865 870 875 880

His Asp Phe Gly Arg Gln Asn Asp Ala Asp Lys Ile Gln Ile Pro Lys 885 890 895

Ile Phe Thr Asn Val Gly Phe Arg Tyr His Leu Glu Ser Pro Ile Ser 900 905 910

Ser Ser Gln Arg Arg Glu Asp Asp Arg Ile Thr Tyr Ile Asn Lys Gly 915 920 925

Gln Phe Tyr Gly Ile Thr Leu Glu Tyr Val His Asp Ala Glu Lys Pro 930 935 940

Ile Lys Asn Thr Thr Val Lys Ser Val Ile Met Leu Met Phe Arg Glu 945 950 955 960

Glu Lys Ser Pro Glu Asp Glu Ile Lys Ala Trp Gln Phe Trp His Ser 965 970 975

Arg Gln His Ser Val Lys Gln Arg Ile Leu Asp Ala Asp Thr Lys Asn 980 985 990

Ser Val Gly Leu Val Gly Cys Ile Glu Glu Val Ser His Asn Ala Ile 995 1000 1005

Ala Val Tyr Trp Asn Pro Leu Glu Ser Ser Ala Lys Ile Asn Ile 1010 1015 1020

Ala Val Gln Cys Leu Ser Thr Asp Phe Ser Ser Gln Lys Gly Gly 1025 1030 1035

Leu Pro Leu His Val Gln Ile Asp Thr Phe Glu Asp Pro Arg Asp 1040 1045 1050

Thr Ala Val Phe His Arg Gly Tyr Cys Gln Ile Lys Val Phe Cys 1055 1060 1065

Asp Lys Gly Ala Glu Arg Lys Thr Arg Asp Glu Glu Arg Arg Ala

1070 1075 Ala Lys Arg Lys Met Thr Ala Thr Gly Arg Lys Leu Asp Glu Leu Tyr His Pro Val Thr Asp Arg Ser Glu Phe Tyr Gly Met Gln - 1110 Asp Phe Ala Lys Pro Pro Val Leu Phe Ser Pro Ala Glu Asp Met Glu Lys Val Gly Gln Leu Gly Ile Gly Ala Ala Thr Gly Met Thr Phe Asn Pro Leu Ser Asn Gly Asn Ser Asn Ser Asn Ser His Ser Ser Leu Gln Ser Phe Tyr Gly His Glu Thr Asp Ser Pro Asp Leu Lys Gly Ala Ser Pro Phe Leu Leu His Gly Gln Lys Val Ala Thr Pro Thr Leu Lys Phe His Asn His Phe Pro Pro Asp Met Gln Thr Asp Lys Lys Asp His Ile Leu Asp Gln Asn Met Leu Thr Ser Thr Pro Leu Thr Asp Phe Gly Pro Pro Met Lys Arg Gly Arg Met Thr Pro Pro Thr Ser Glu Arg Val Met Leu Tyr Val Arg Gln Glu Asn Glu Glu Val Tyr Thr Pro Leu His Val Val Pro Pro Thr Thr Ile Gly Leu Leu Asn Ala Ile Glu Asn Lys Tyr Lys Ile Ser Thr Thr Ser Ile Asn Asn Ile Tyr Arg Thr Asn Lys Lys Gly Ile Thr Ala - 74 -

1280 1285 1290

Lys Ile Asp Asp Asp Met Ile Ser Phe Tyr Cys Asn Glu Asp Ile 1295 1300 1305

Phe Leu Leu Glu Val Gln Gln Ile Glu Asp Asp Leu Tyr Asp Val 1310 1315 1320

Thr Leu Thr Glu Leu Pro Asn Gln 1325 1330

**DATED** this twenty-second day of August 2002.

## Melbourne Health

by DAVIES COLLISION CAVE

Patent Attorneys for the Applicant

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